

Blood Based Cell Biopsy For Cancer Diagnostics



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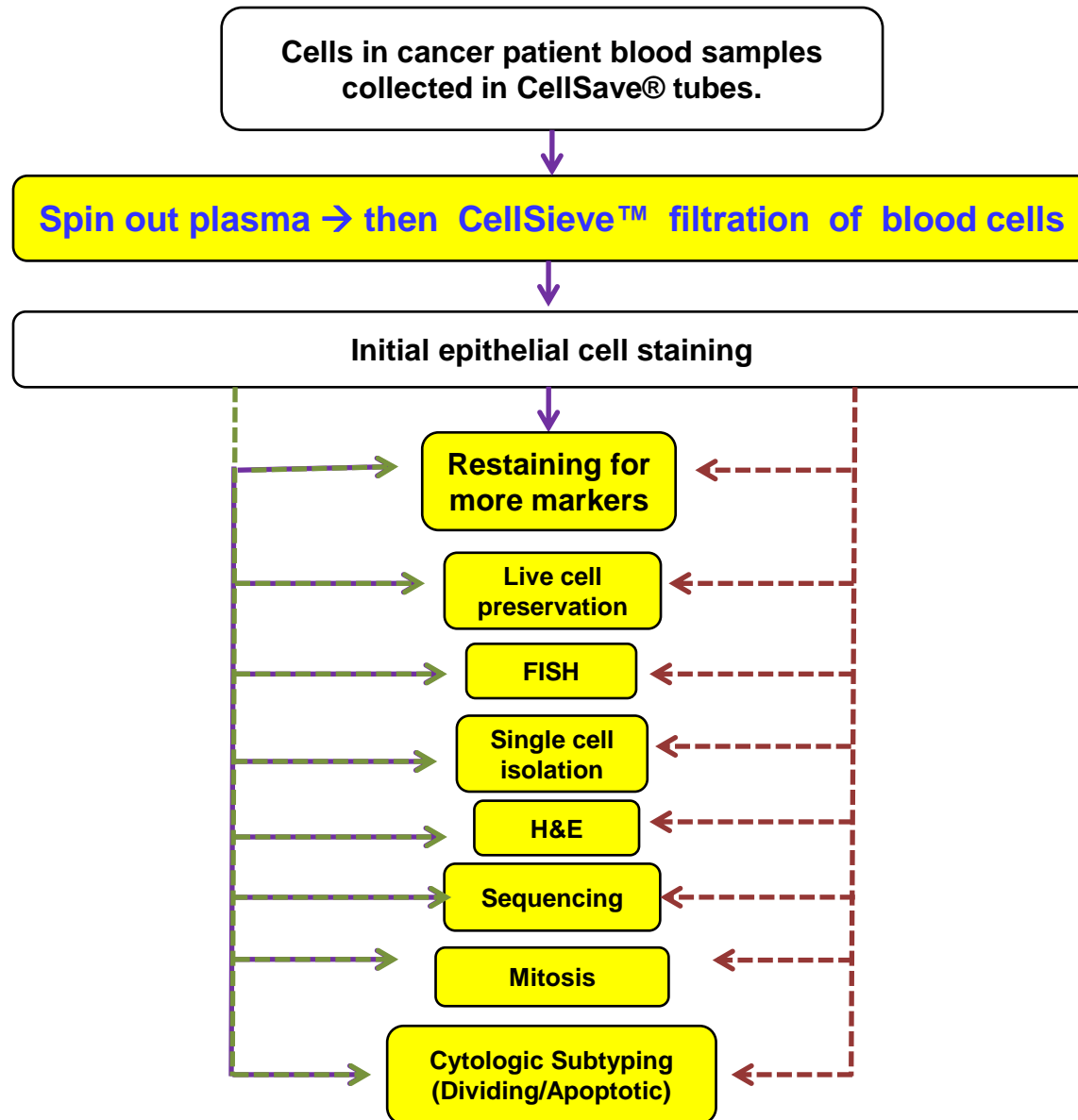
Outline

- **Overview**
- **Technical background**
- **Clinical application examples**

Unmet Needs

- **Blood based companion diagnostics**
 - **example immunotherapy**
- **Monitor treatment response**
- **Early detection of cancer**
- **Early detection of recurrence**
- **R&D and drug development applications**

Overview of Assays Capabilities

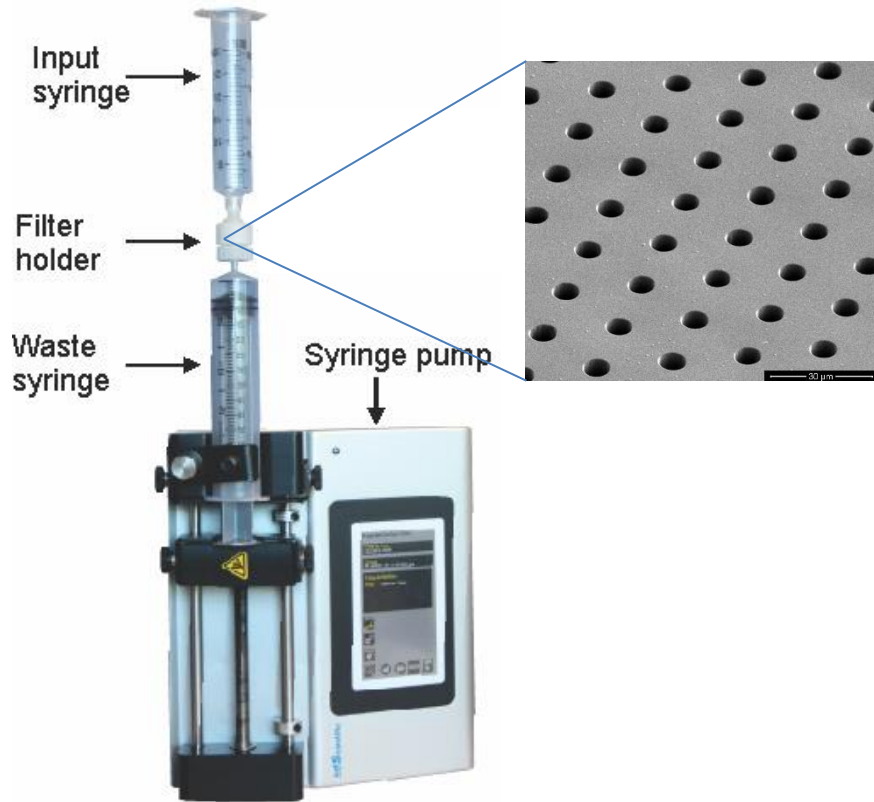


Clinical Sample Type

- Peripheral blood
- Bone marrow
- Cryo preserved samples
- Urine

CellSieve™ Microfilters

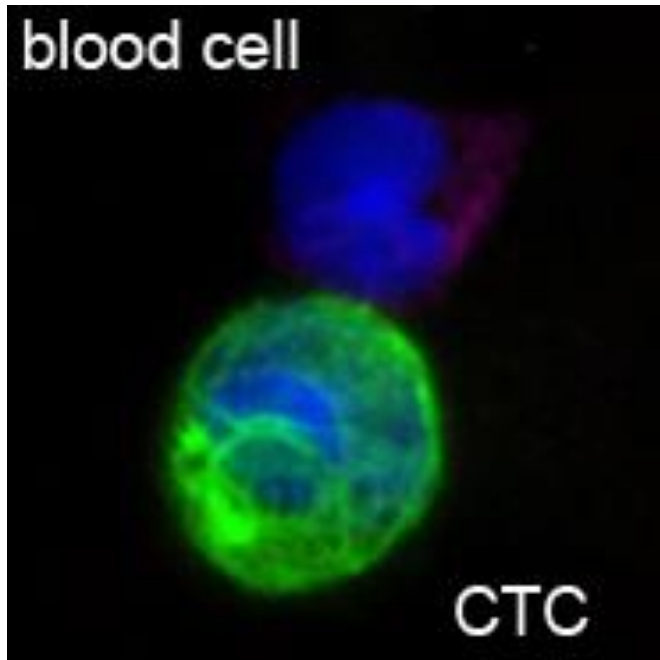
Not based on surface markers



- **Uniform 7 µm pore size and distribution with high porosity**
 - Rapid, consistent and gentle flow
 - 3 min to filter 7.5 ml of blood
 - Eliminates all red blood cells
 - Eliminates ~99.99% of white blood cells
- **Low fluorescence background**
- **CellSave tubes good ≤ 96 hrs**

“Pathologically Defined” CTCs

Creatv's criteria to improve accuracy

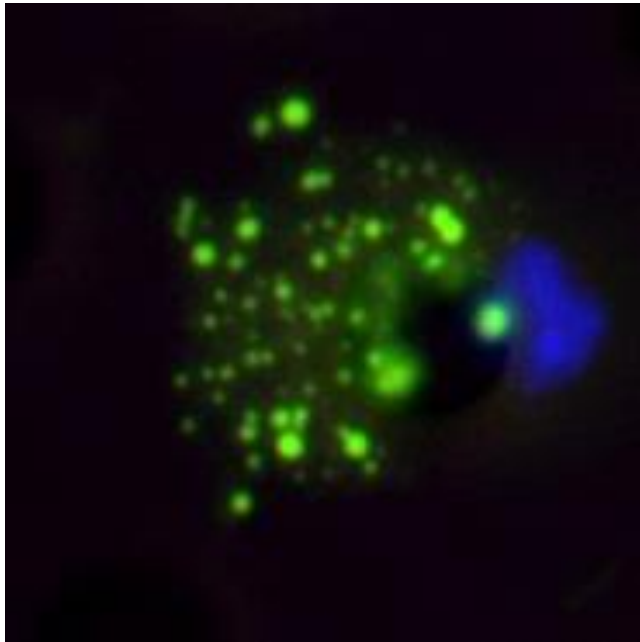


- DAPI positive
 - and cancerous looking
- To identify CTC
 - CK 8, 18, 19 (+)
 - Cytoskeletons → filamentous pattern
- To rule out white blood cells
 - CD45 (-)

Feasible with CellSieve™ microfiltration system

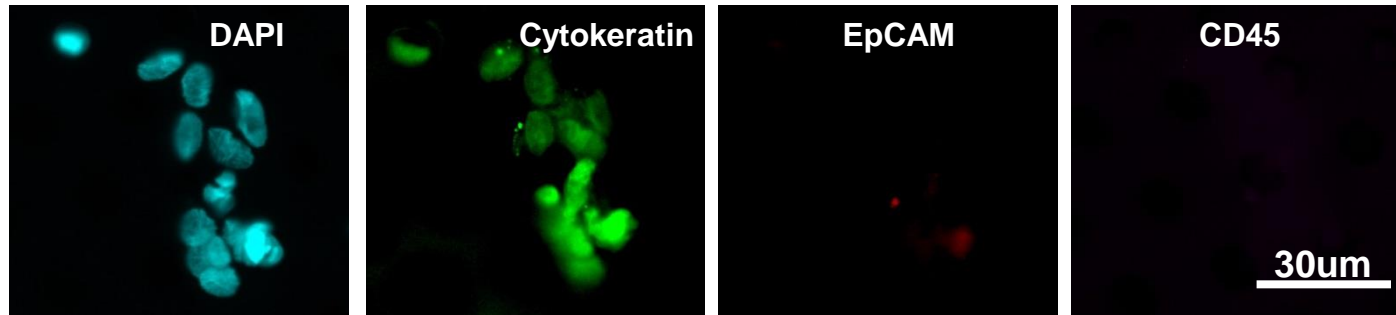
Apoptotic CTCs

Creatv's criteria to improve specificity



- DAPI positive
 - and nucleus degrading
- CK 8, 18, 19 (+)
 - and fragmented into **spots**
- To rule out white blood cells
 - CD45 (-)

Epithelial Mesenchymal Transition Cells (EMTs) – cells can later become cancer cells

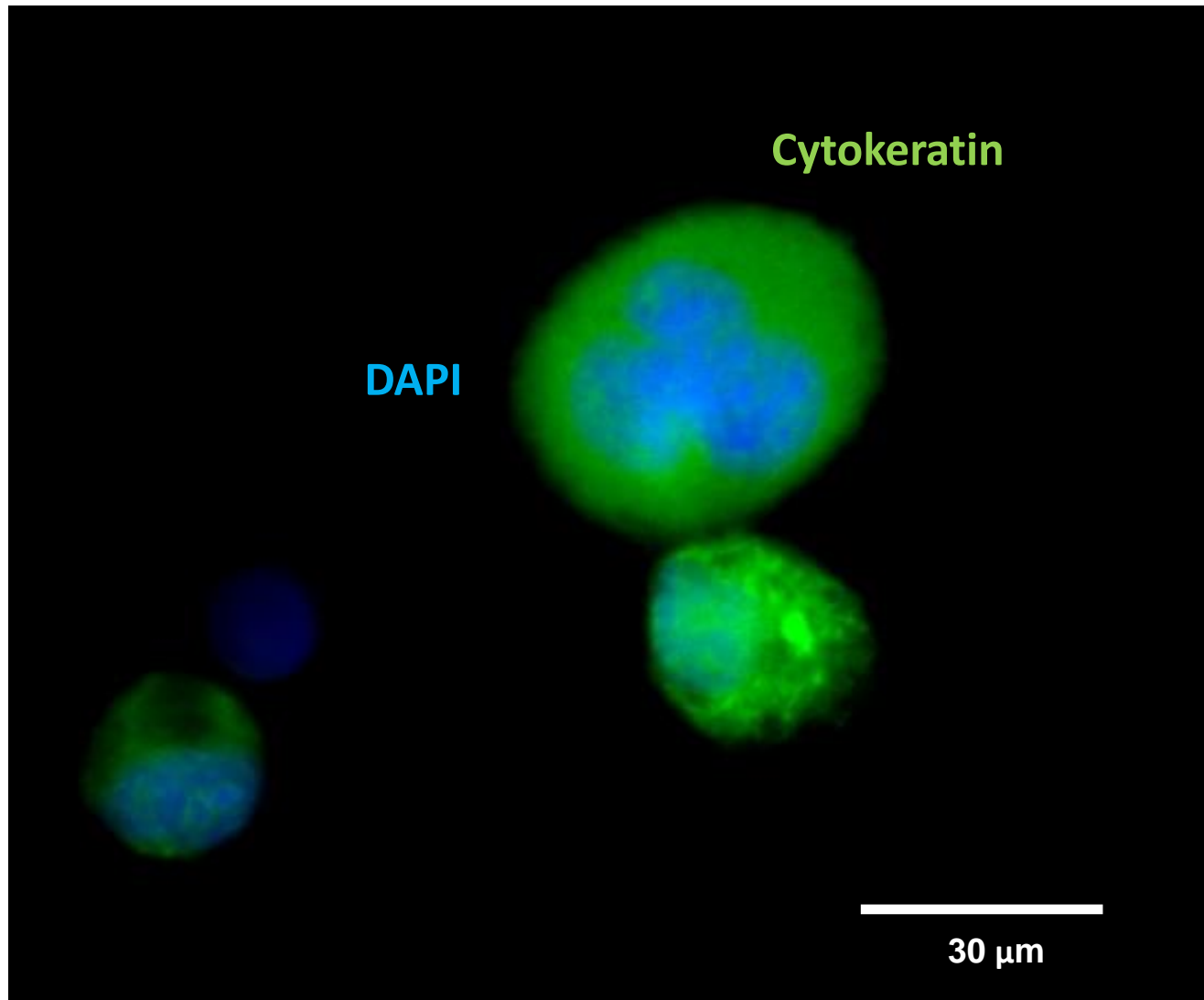


- DAPI positive
 - Smaller nuclei than CTCs
 - Usually form clusters
- Identification of EMTs
 - Weakly CK 8, 18, 19 (+)
 - No EpCAM
- To rule out white blood cells
 - CD45 (-)

Creatv Discovers a New Cell Type

Cytokeratin: marker for epithelial cell

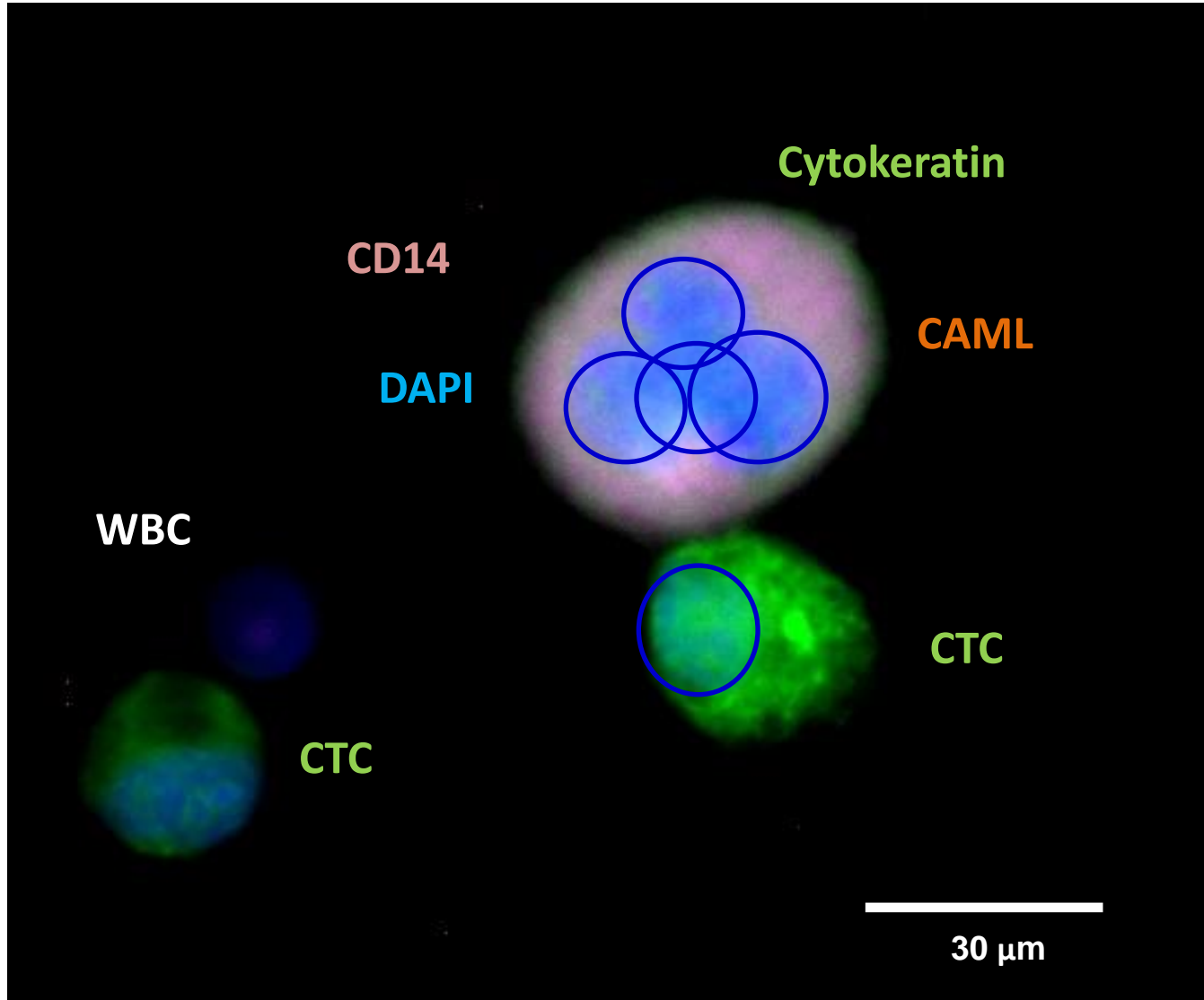
DAPI: marker for the nucleus



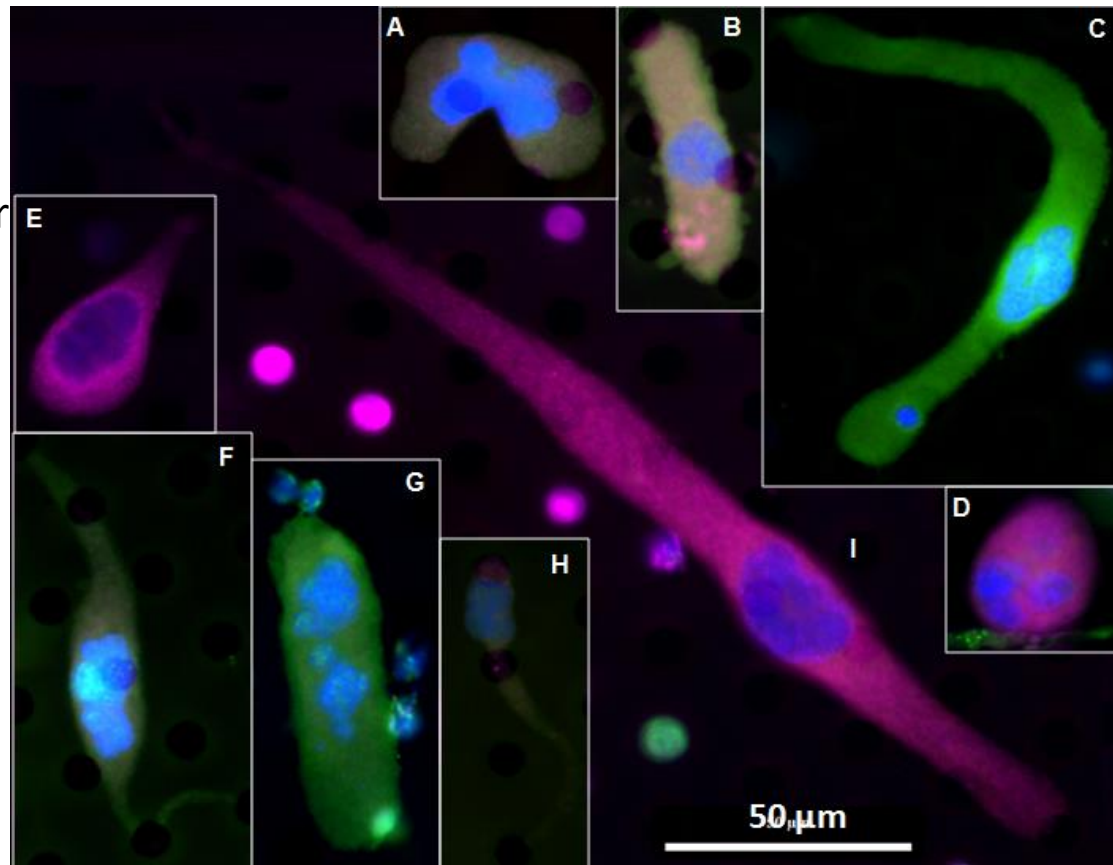
Discovery

CTC: circulating tumor cells

CAML: circulating cancer associated macrophage-like cells

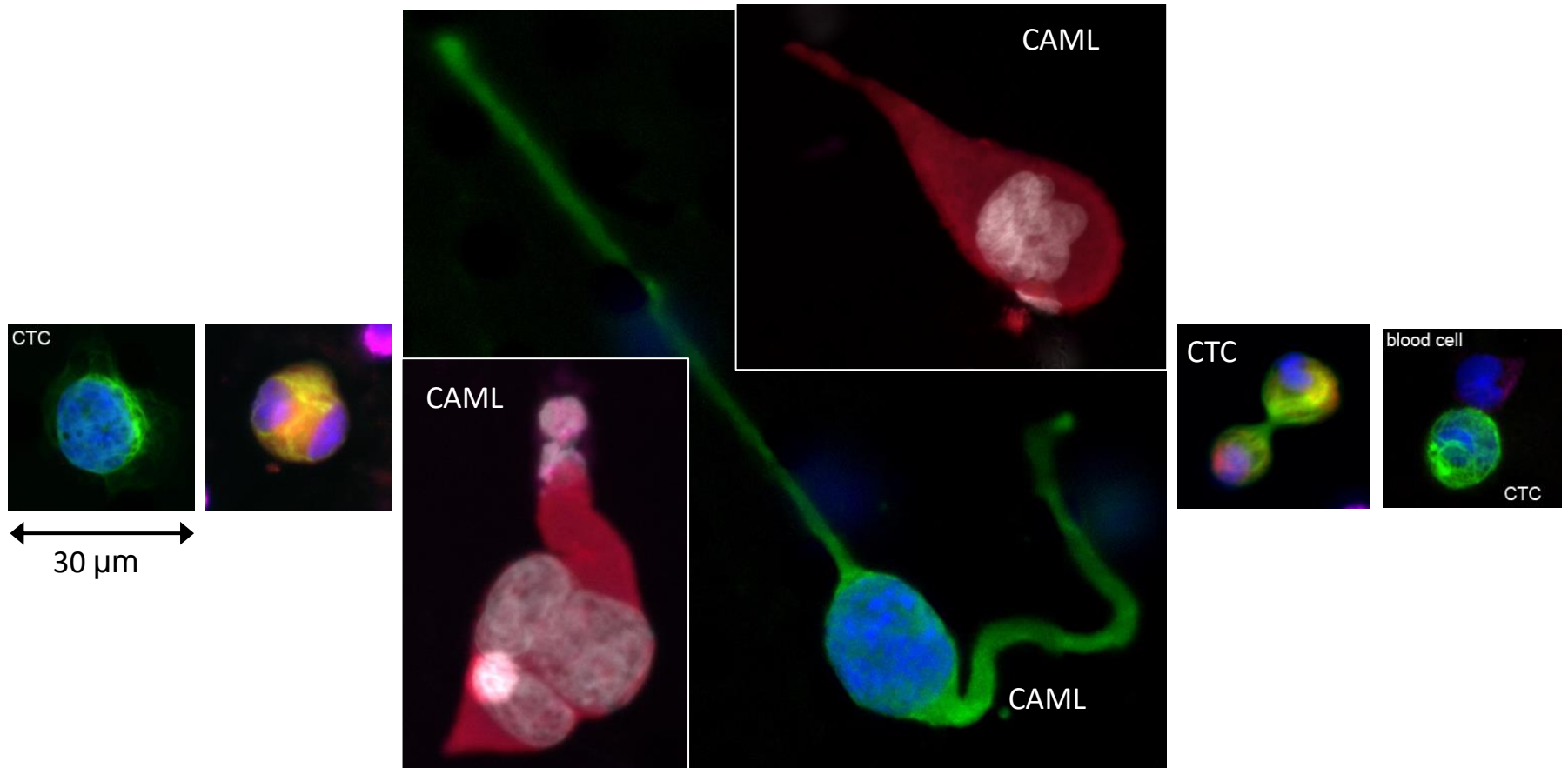


Circulating Cancer Associated Macrophage-like Cells (CAMLs)



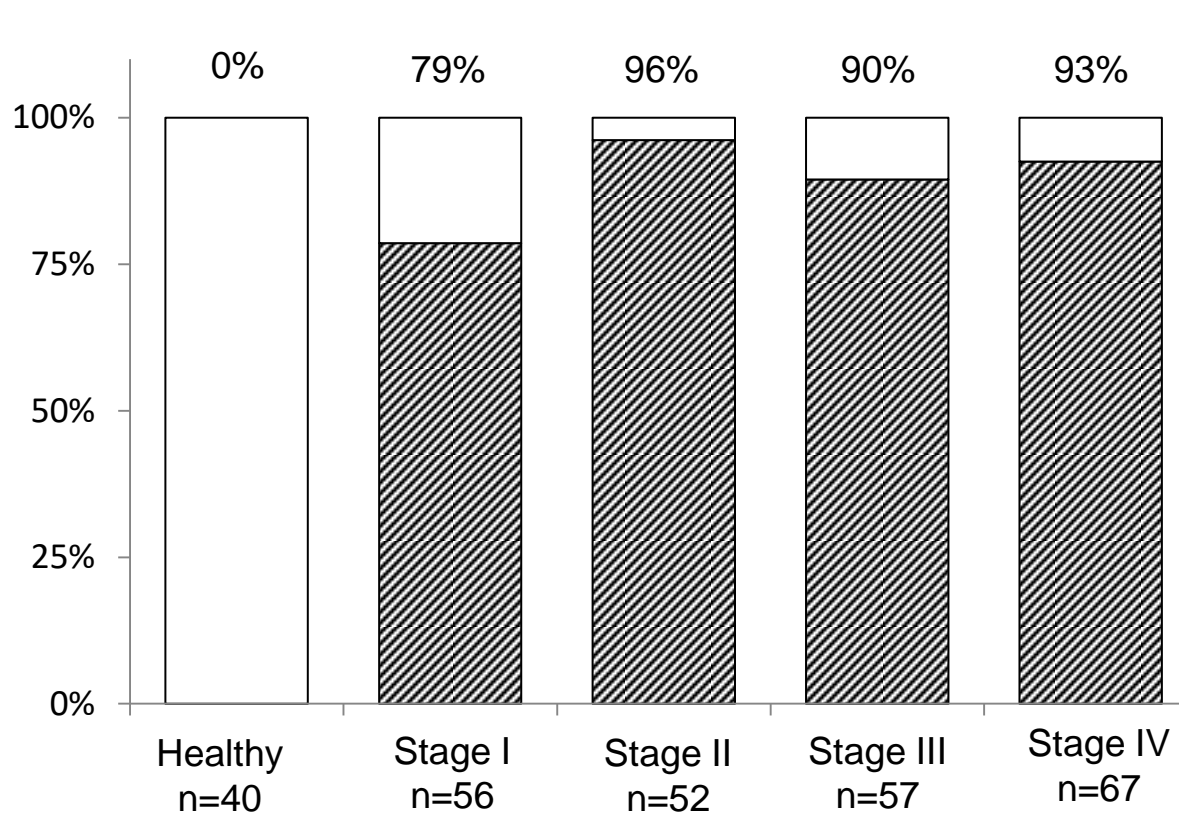
- Large, atypical nucleus
- May express CK and EpCAM
- Contain tumor markers
- Most are CD45 positive
- Large: 25 - 300 μm
- Express CD11c/CD14
- Express endothelial markers CD146, TIE-2

CAMLs Engulf Tumor Cells and Debris



CAMLs only occur in Cancer Patients

None in healthy controls



Total
n=272*

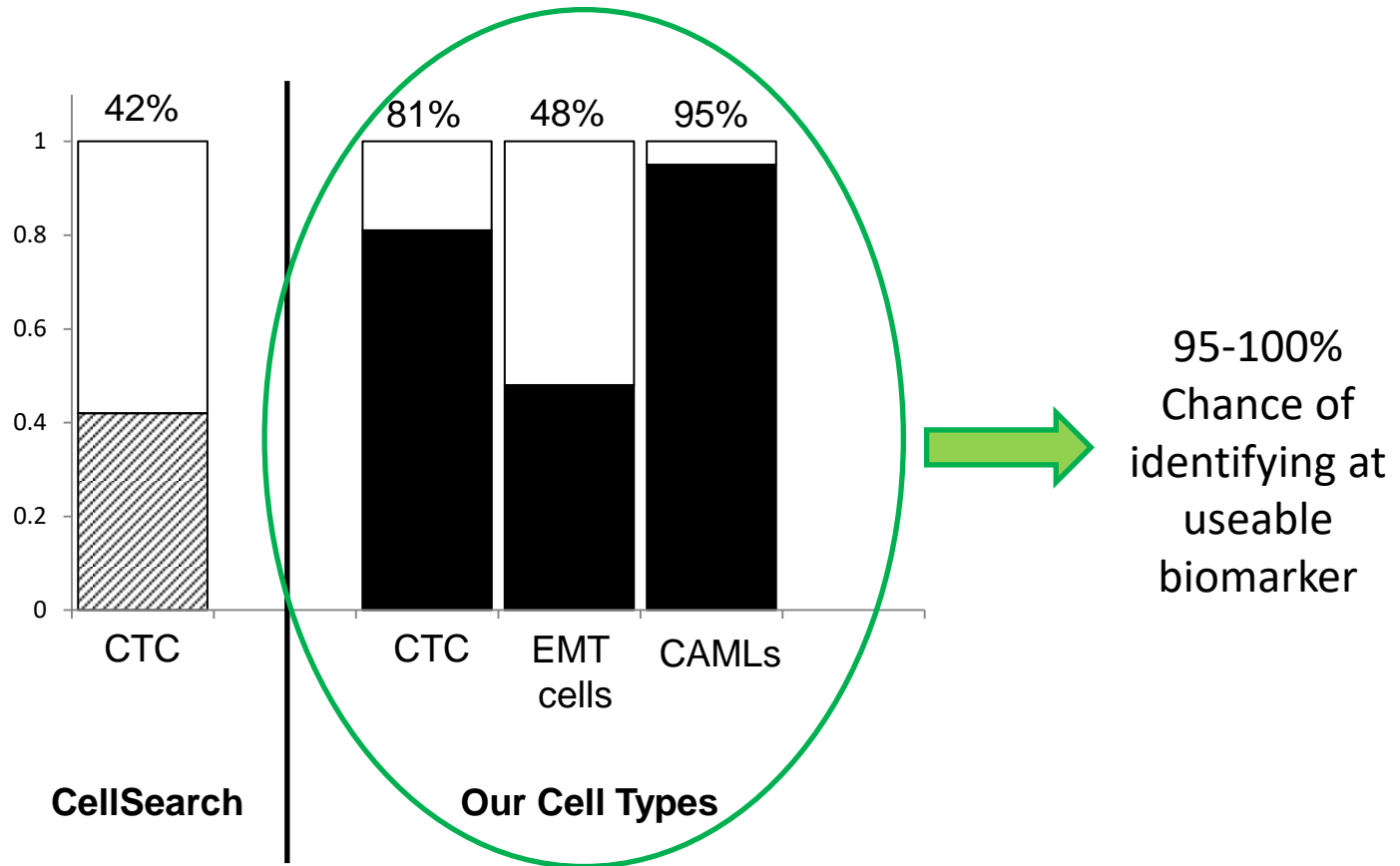
Cancer types

- Breast
- Prostate
- Pancreatic
- Lung (NSCLC)
- Colon
- Esophageal

Sensitivity 89% (95% CI 85-93%)
Specificity 100% (95% CI 91-100%)
PPV 100% (95% CI 98-100%)

We analyze all cancer-associated cells to maximize useable biomarkers

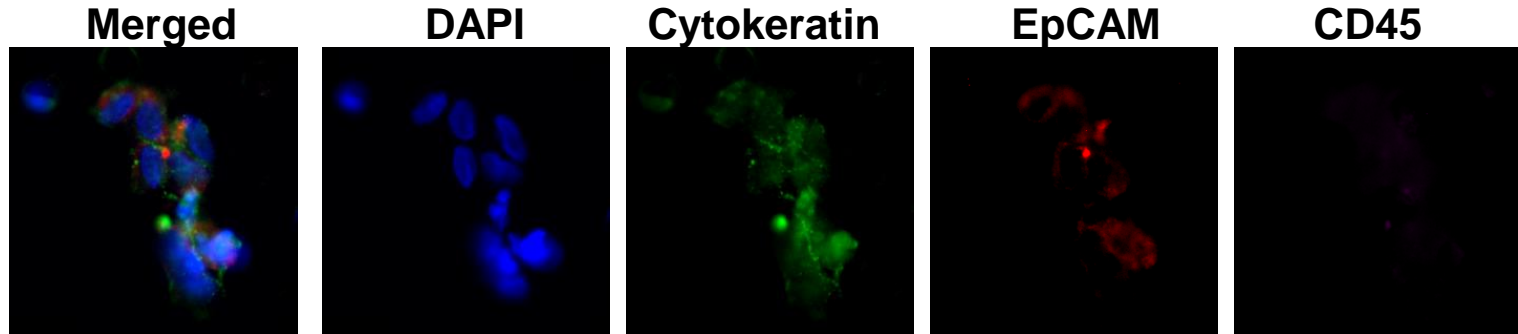
Presence of cell types in **Breast Cancer Patients**



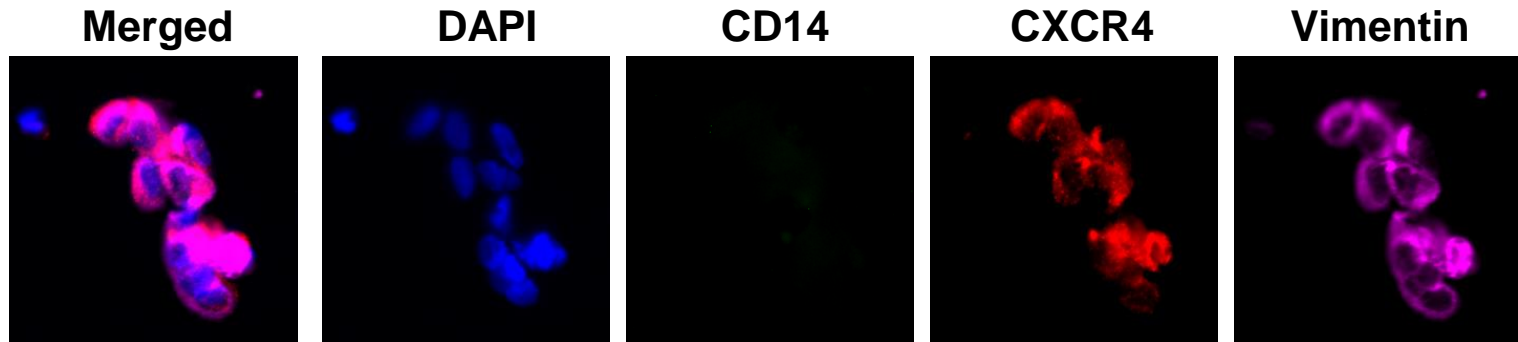
- **Companion diagnostics utilizing CAMLs**
 - Evaluate drug targets – not limited by availability of tissue
- **Monitor treatment response**
 - CAMLs and CTCs are independent indicators
 - Real-time measure of response
- **Clinical Trials**
 - Cohort selection
 - Real-time measure of response – even for immunotherapy
- **Cancer Screening**
 - At-risk populations: Lung, pancreatic, colorectal, etc.
 - Hard-to-detect cancers
 - Cancer recurrences

Multi-analyte Subtyping Restaining Technique

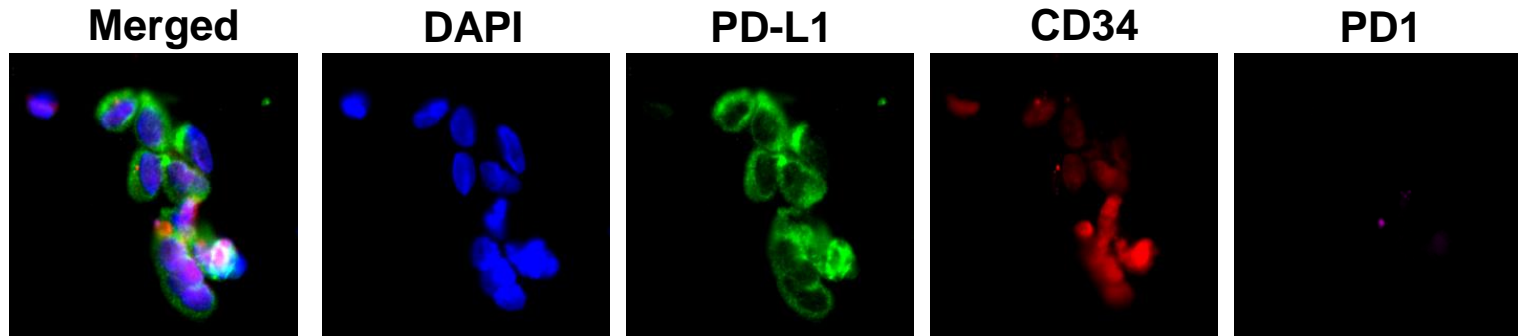
Initial CTC
stains



Subtyping
cells



Therapy
targeting



Immunotherapy Diagnostic Needs

■ Companion diagnostics

- Currently based on tissue biopsy
- Biopsy not able to guarantee inclusion of stromal infiltrated regions
- Usually not repeatable

■ Monitor treatment

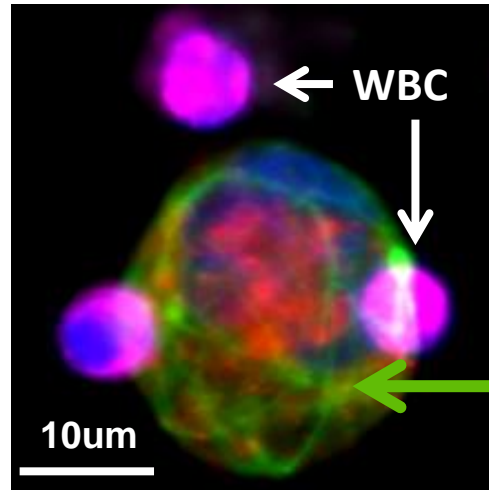
- Imaging cannot monitor treatment response because stromal infiltration causes the tumor to become larger (pseudo-progression)

CellSieve™ Analysis of Immunotherapy

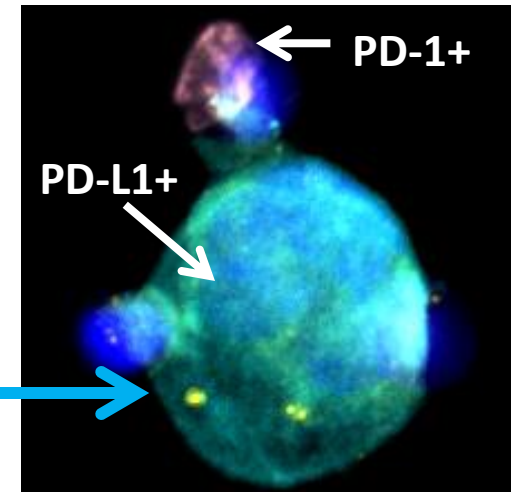
Breast

CTC with bound
white blood cells

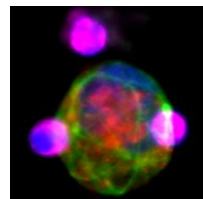
Nucleus(dark blue)/CK(green)/
EpCAM(red)/CD45(violet)



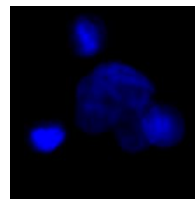
Nucleus(dark blue)/PD-L1(turquoise)/
PD-1(pink)



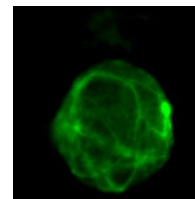
Merged



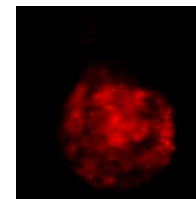
DAPI



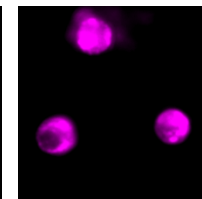
Cytokeratin



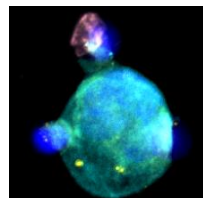
EpCAM



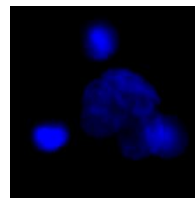
CD45



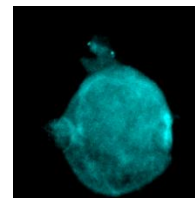
Merged



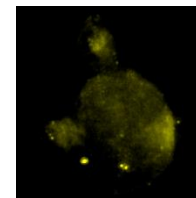
DAPI



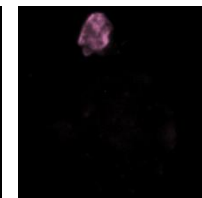
PD-L1



x



PD-1

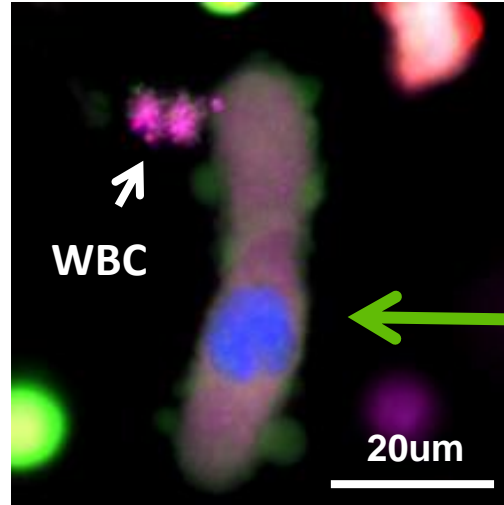


CellSieve™ Analysis of Immunotherapy

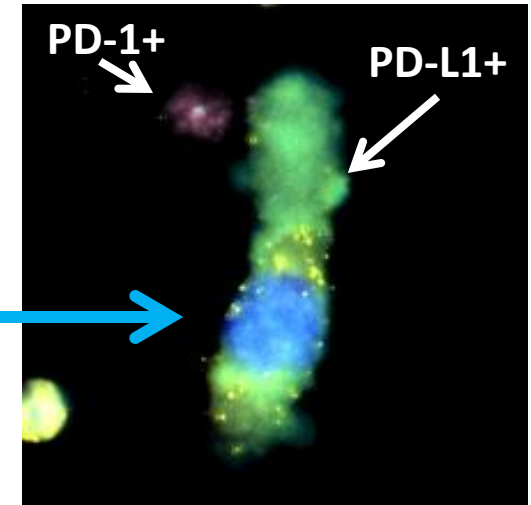
CAML

Cytokeratin positive
cell with bound
white blood cell

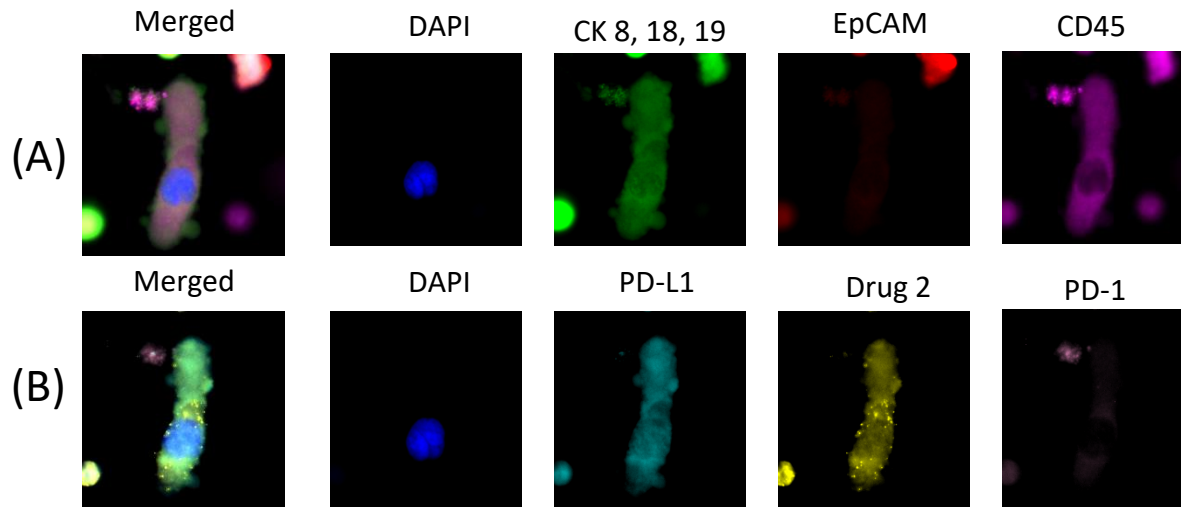
Nucleus(dark blue)/CK(green)/
EpCAM(red)/CD45(violet)



Nucleus(dark blue)/PD-L1(turquoise)/
PD-1(pink)



CAML



CTCs and CStCs

identify the upregulation or down regulation of immunotherapy in real time

- **Oral Presentation WCLC 2015** - Sequential Assessment of DNA damage response and PD-L1 expression in circulating tumor cells of lung cancer patients during treatment with radiotherapy
- **Poster Presentation AACR 2016** - Sequential tracking of PD-L1 expression and RAD50 induction in CTCs and circulating stromal cells of lung cancer patients during treatment with radiotherapy

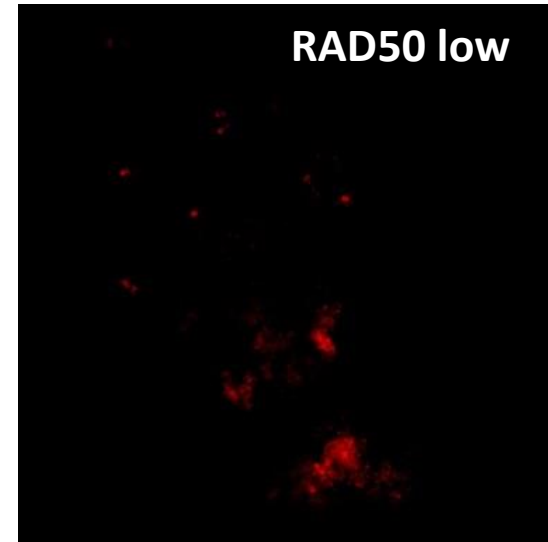
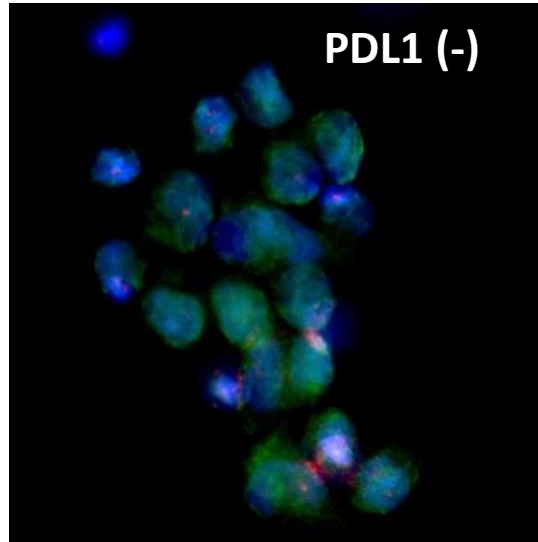
Effect of RAD50 and PDL1

NSCLC – before and after radiation therapy

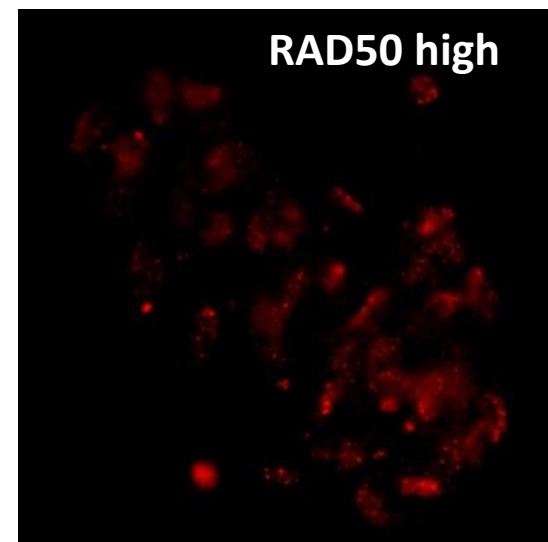
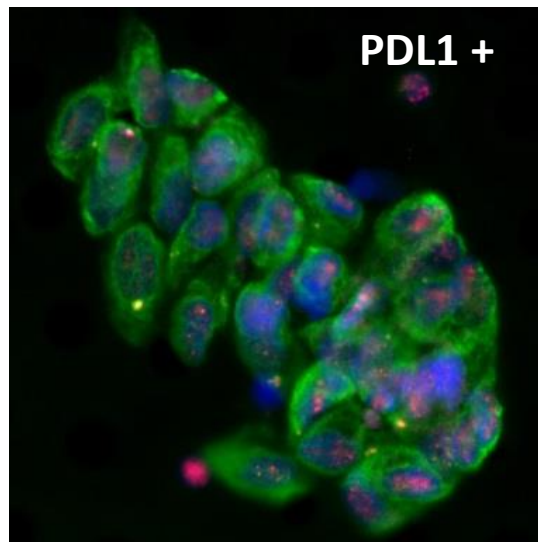
Merge

RAD50

Before
Radiation

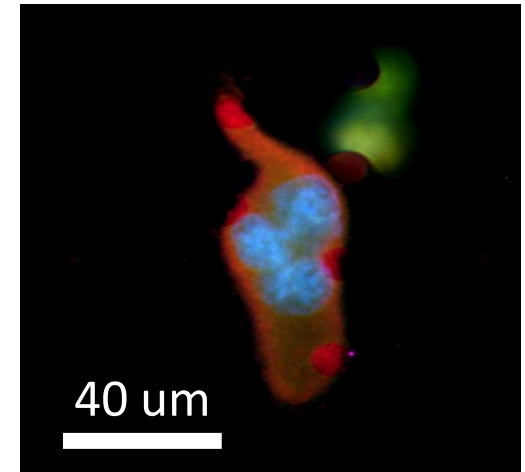


Post
Radiation



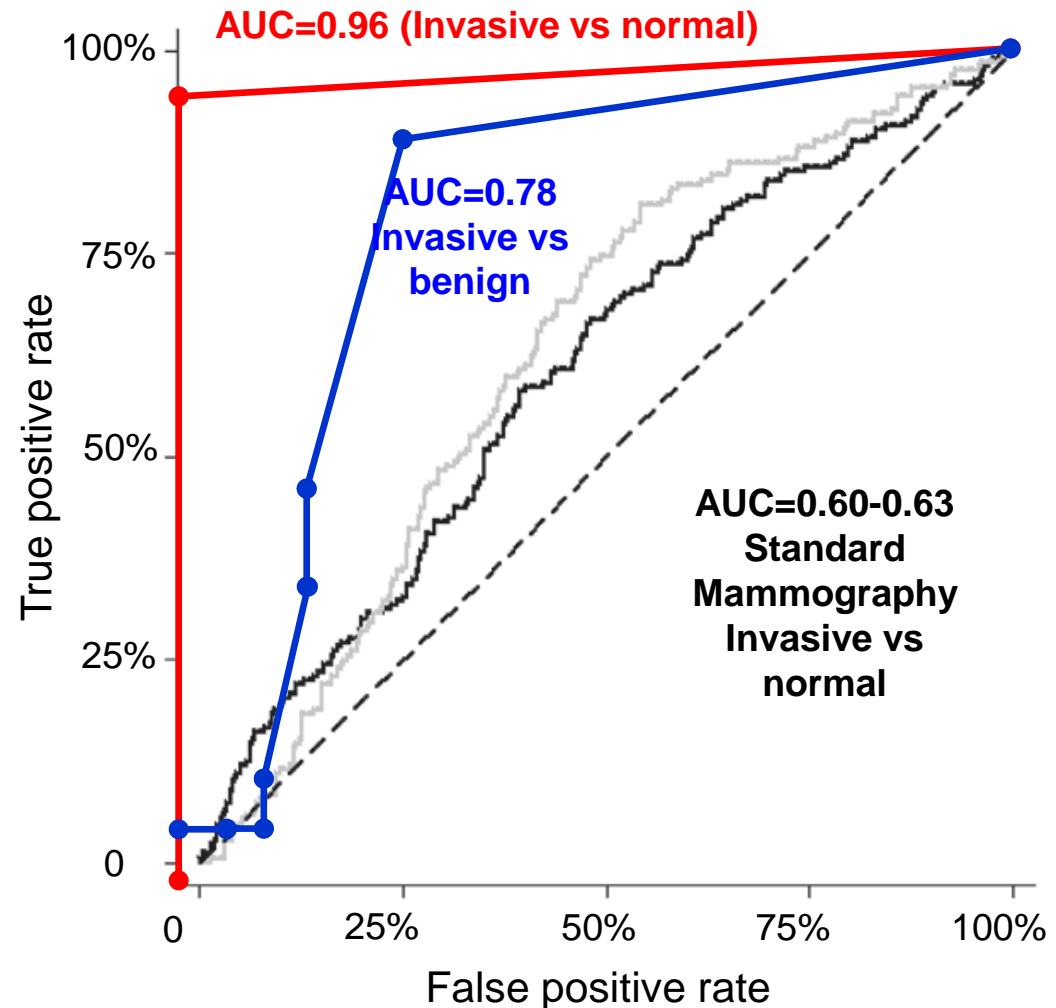
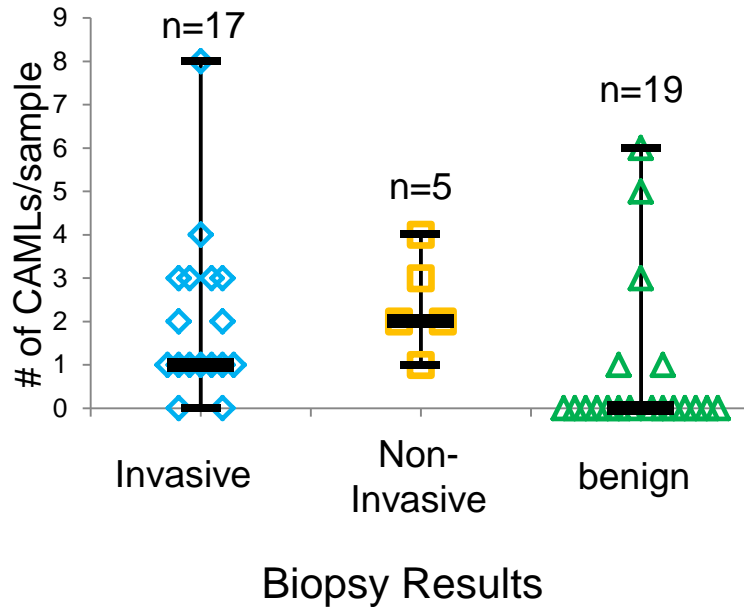
Double blinded study

- **41 mammography positive patients**
 - Median age 52
- **Double blinded study**
 - Tissue biopsy
 - CAML blood testing
- **Blood test criteria – 1 or more CAML**
 - $\geq 30 \mu\text{m}$
 - CD14 (+)
- **CAMLs differentiated benign vs malignant conditions**
 - Sensitivity (+ for cancer) – 88%
 - Specificity (no cancer) – 74%



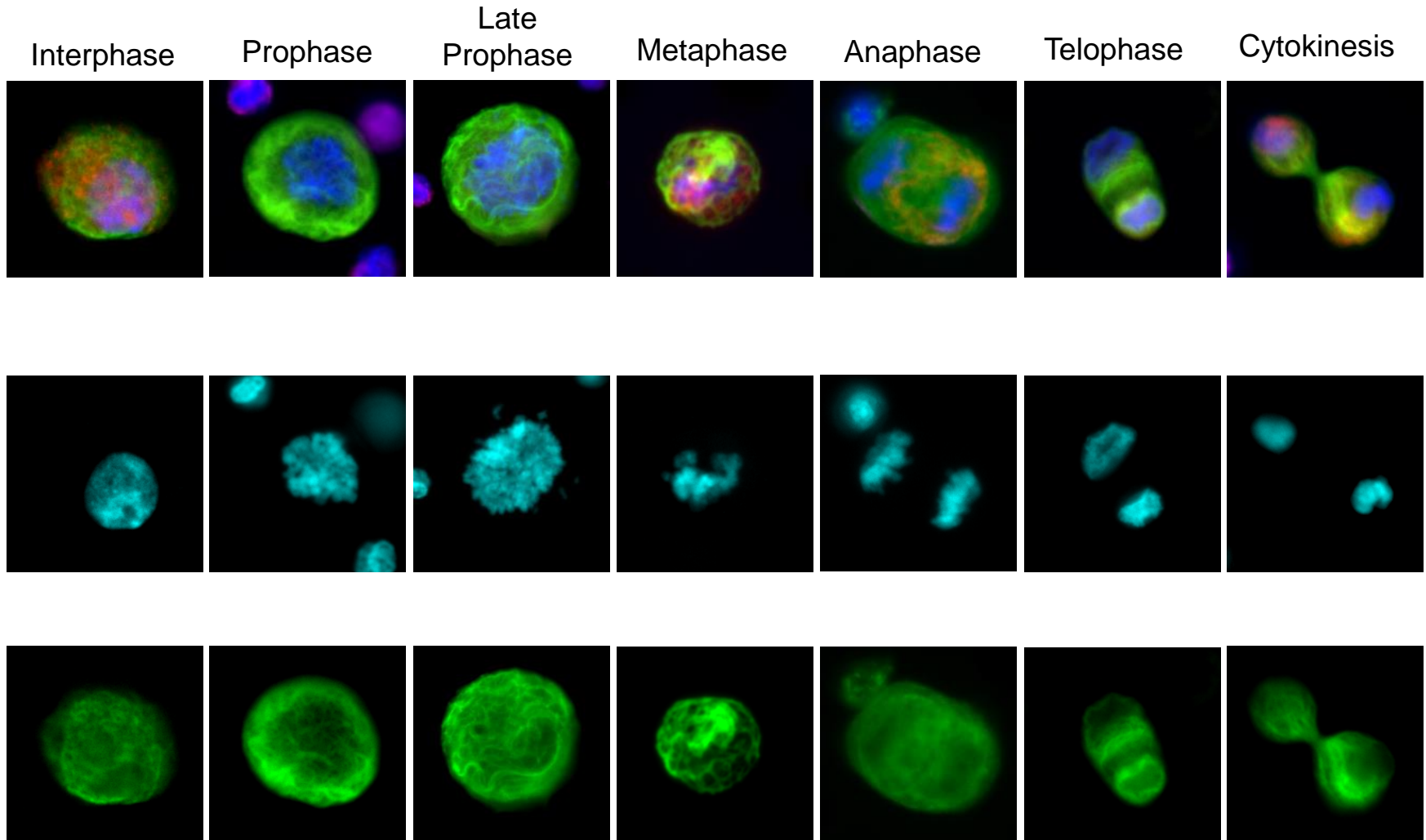
CAMLs for Breast Cancer Screening

Superior to mammography



CTCs in Division

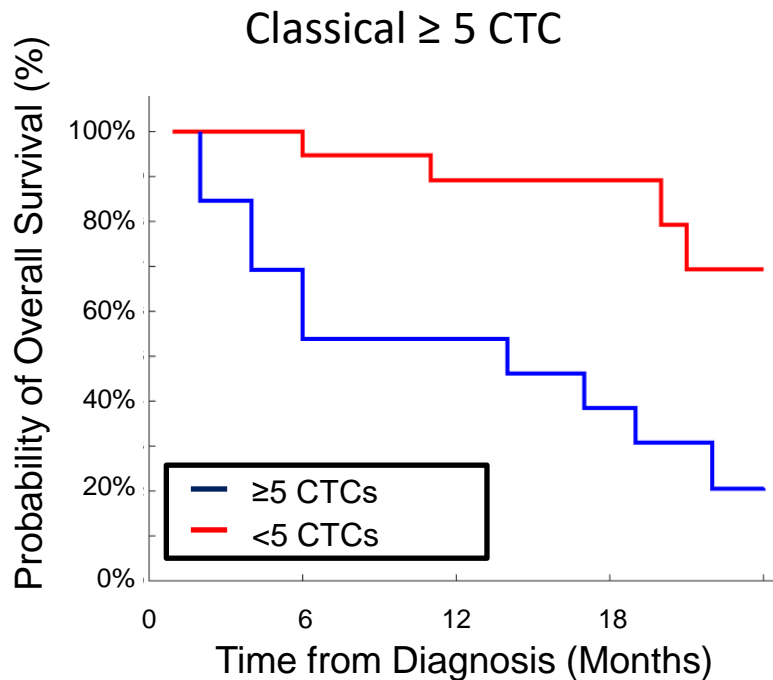
Cytokeratin (green), DAPI(blue)



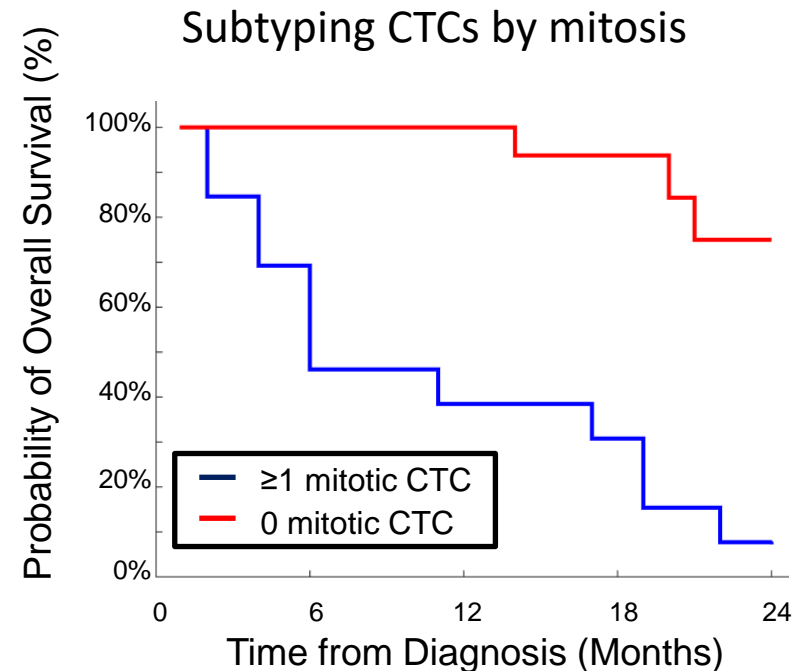
CTCs subtyping by mitotic events

Increased prognostic value than CTC count

Greater or less than 5 CTCs **vs** with and without a mitotic CTC event
(n=36)



Hazard Ratio: 5.2



Hazard Ratio: 11.1

Summary

- **CellSieve™ Liquid Cell Biopsy – isolates CTCs and stromal cells**
 - Applicable to many types of assays
 - Rapid, straightforward workflow
 - High sensitivity
 - High specificity
- **Very useful biomarker – CAML**
 - Found in all stages of cancer
 - Express cancer markers
 - Very easy to identify
 - More prevalent than CTCs
- **Same blood sample can be used for cfDNA**
- **Wide variety of assays and clinical utility**

Research Collaborators

Research Institute	Collaborators
University of Maryland Baltimore	Stuart Martin, Ph.D., Monica Charpentier, M.D. Martin Edelman, M.D., Rena Lepidus, Ph.D.
Northwestern University	Massimo Cristofanilli, M.D.
Fox Chase Cancer Center	R. Katherine Alpaugh, Ph.D.
Johns Hopkins University	David Loeb, M.D.
Mayo Clinic Cancer Center	Thai Ho, M.D., Saranya Chumsri, M.D.
MD Anderson	Steven Lin, M.D.
Medical College of Wisconsin	Susan Tsai, M.D.
OHSU Knight Cancer Institute	Raymond C. Bergan, M.D.
Duke University	Jeffery Marks, Ph.D.
Memorial Sloan Kettering Cancer Center	Daniel Danila, M.D.
Washington University	Rebecca Aft, M.D.
University of Chicago	Susan Cohn, M.D.
George Washington University	Christian C. Haudenschild, M.D.
Hememics Biotechnologies	Steigrimur Stefansson, Ph.D.

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The content of the information does not necessarily reflect the position or the policy of the US Government.



Thank you

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