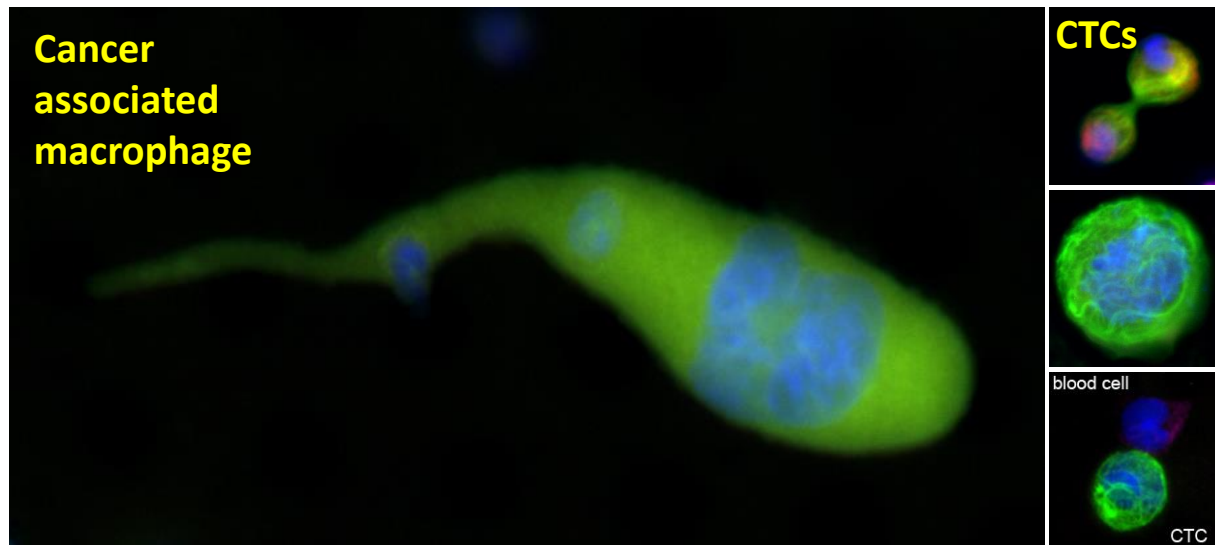


Circulating Stromal Cells in Immunotherapy

Utilizing a Total Blood Based Biopsy



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Senior Research Scientist/Head of Clinical Core Laboratory

Creatv MicroTech, Inc.

August 24, 2016



Disclosures

- **Employee of Creatv MicroTech, Inc.**
- **Multiple patents on the technologies discussed**

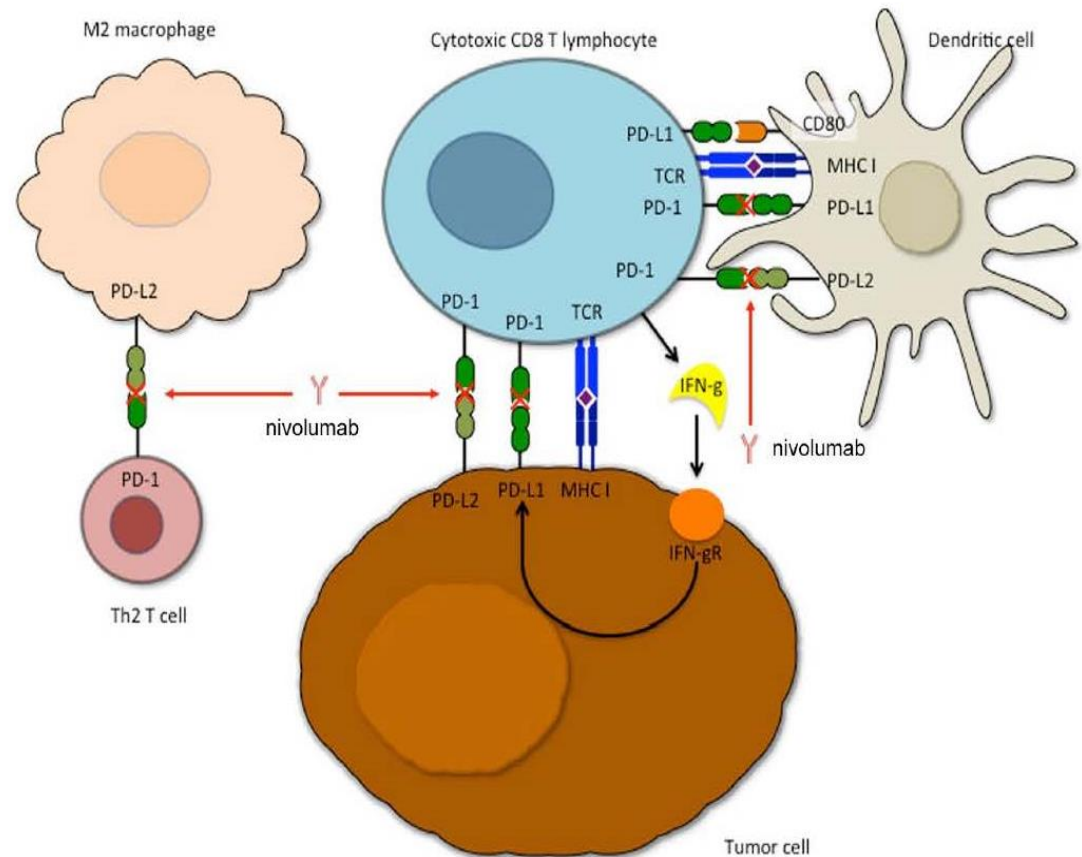
Utilities of Blood Based Biopsy for cancer diagnostics

- **Circulating Tumor Cells (CTCs) if present**
 - Sequential tracking of therapy
 - Genomic and proteomic profiling of tumor
- **CTCs alone not enough and additional needs**
 - EARLY and late stage diseases (CTCs only late)
 - Majority of patient population (CTC only certain pop.)
 - Immunotherapy (targets non-malignant cells)
 - Assist drug development

Immunotherapies involve multiple cell types (Ex: PD-1/PD-L1 pathway)

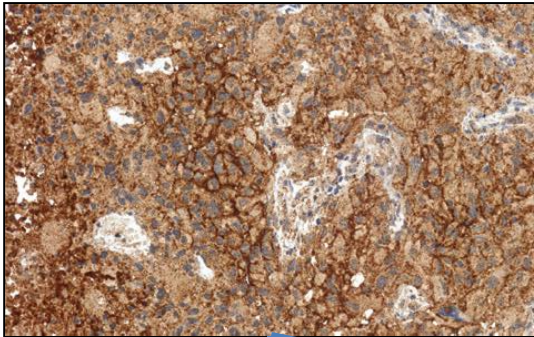
PD-L1 can be found on:

- Tumor cells
- Stromal macrophages
- Stromal Tc cells
- Stromal Th cells
- Stromal Dendritic cells
- Tumor fibroblasts
- Others

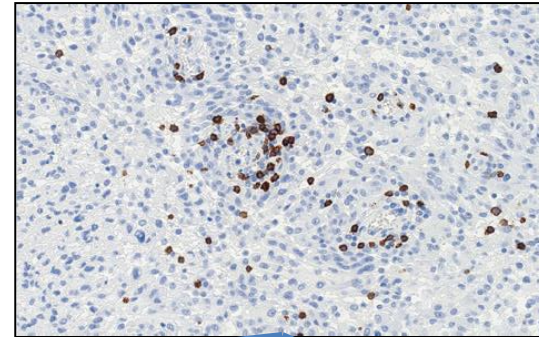


We must rethink how companion diagnostics work

Biopsy tumor cells



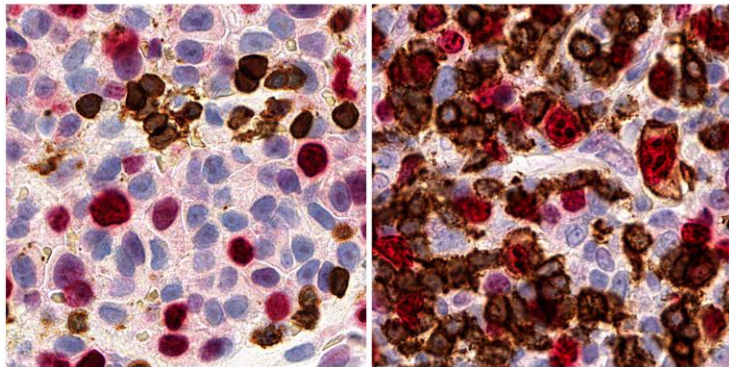
Analyze Immune cells



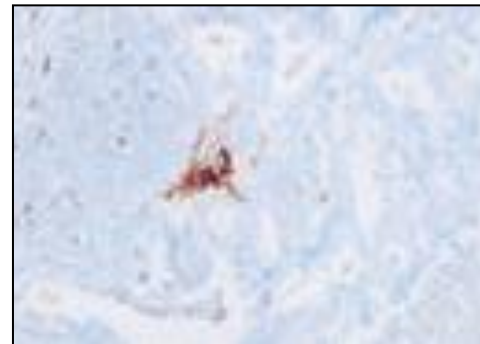
Response

Before Treatment

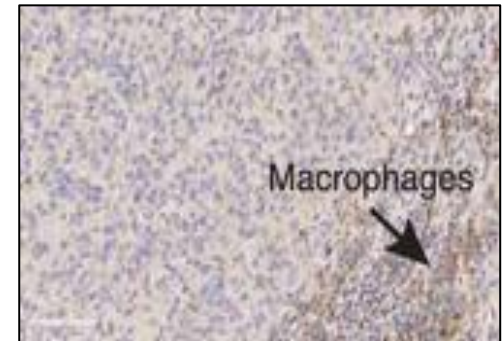
After Treatment



Analyze heterogeneity



Does biopsy contain stroma?



Immunotherapeutic biomarkers must have multi-cellular capabilities

- **Circulating Tumor cells (CTCs)**
- **Circulating Stromal cells (CStCs)**
 - Tumor derived endothelial cells
 - Epithelial-mesenchymal transition cells (EMTs)
 - Tumor associated macrophage-like cells (CAMLs)
 - Tumor derived T cells
 - Tumor associated fibroblasts

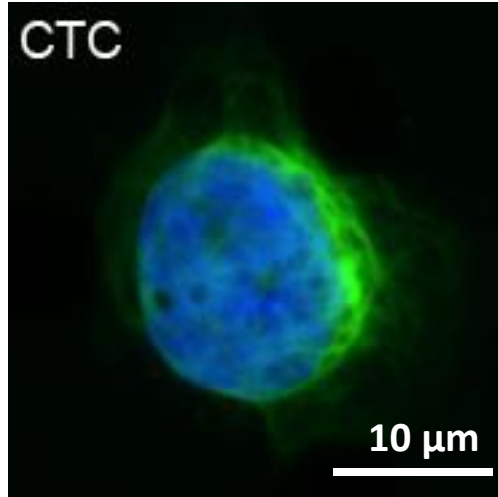
Requirements of Blood Based Biopsy for Immunotherapy

- **Applicable to sequential tracking of therapy**
- **Allow genomic and proteomic profiling of tumor**
- **Applicable to all stages of disease**
- **Must isolate heterogeneous populations of **Multiple cell types****
- **Must isolate heterogeneous populations of **stromal cells**** (Targeting non-malignant cells)

Advantages of Circulating Tumor Cells (CTCs)

■ Advantages

- Provides prognostic information
- Tracks response to therapy
- May provide:
 - Genomic profiling of tumor/metastases
 - Proteomic profiling of the tumor/metastases



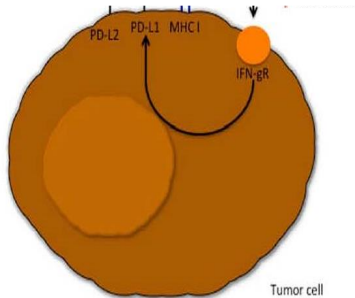
Pathologically defined CTCs (PDCTC)

- CK 8, 18, 19 (+) and filamentous
- DAPI (+) cancerous morphology or in division
- CD45 (-)

Disadvantages of CTCs

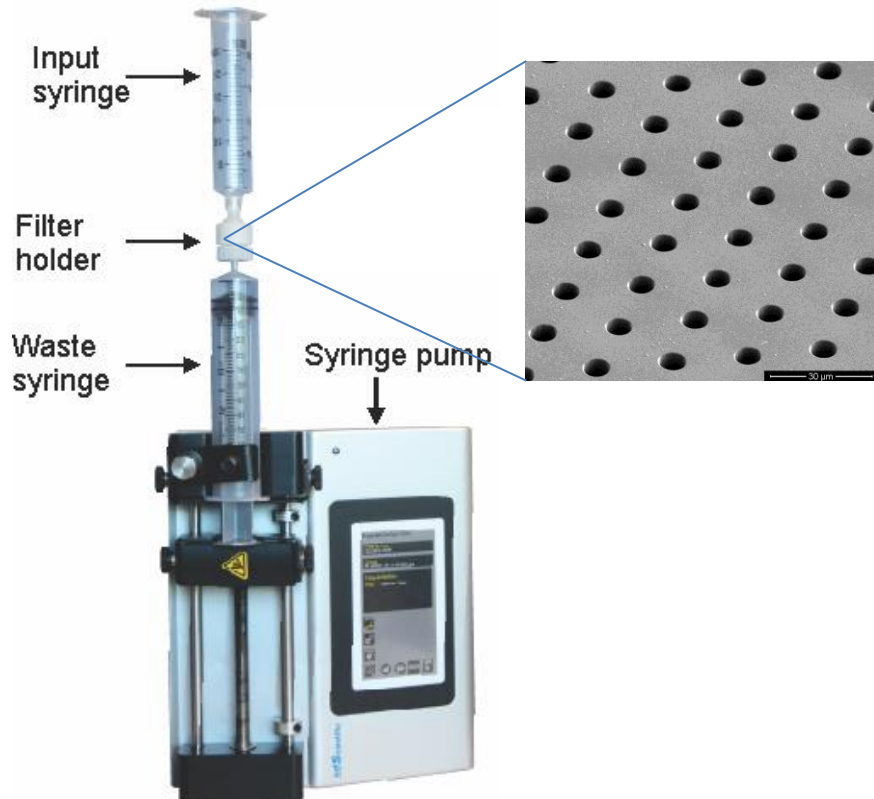
■ Disadvantages

- **Uncommon** (~0-10 per mL blood)
- **Low frequency** (19%-57% of malignant carcinomas)
- Only found in late stage/metastatic
- Tumor cells alone do not represent the stromal environment



Cell Isolation Based on Size

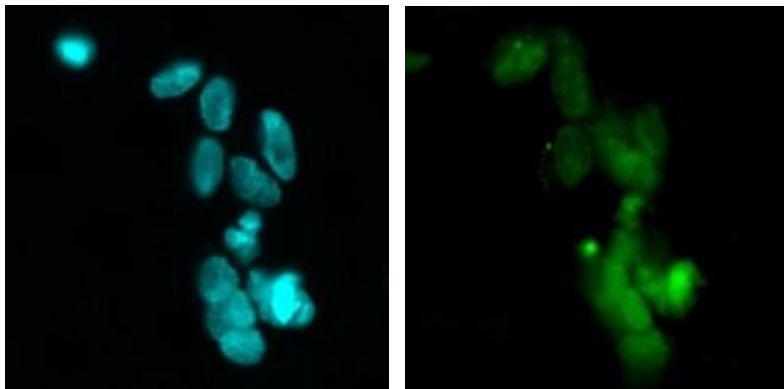
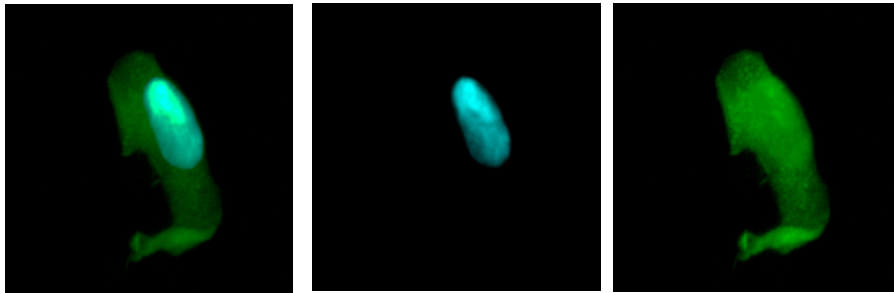
CellSieve™ Microfilters



- **Uniform 7 µm pore size and distribution with high porosity**
 - Rapid, consistent and gentle flow
 - 3 min to filter 7.5 ml of blood
 - Small (100µL) and large (>30mL) sample size
- **Non-fluorescence**
- **CellSave tubes are run ≤ 96 hrs**

EMT like Cells

EMT like Cells



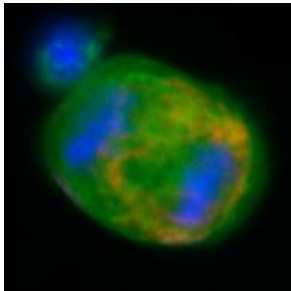
Criteria unique to high resolution imagery

- CK 8, 18, 19 (+) diffuse/non-filamentous
- DAPI (+) cancerous morphology
- CD45 (-)

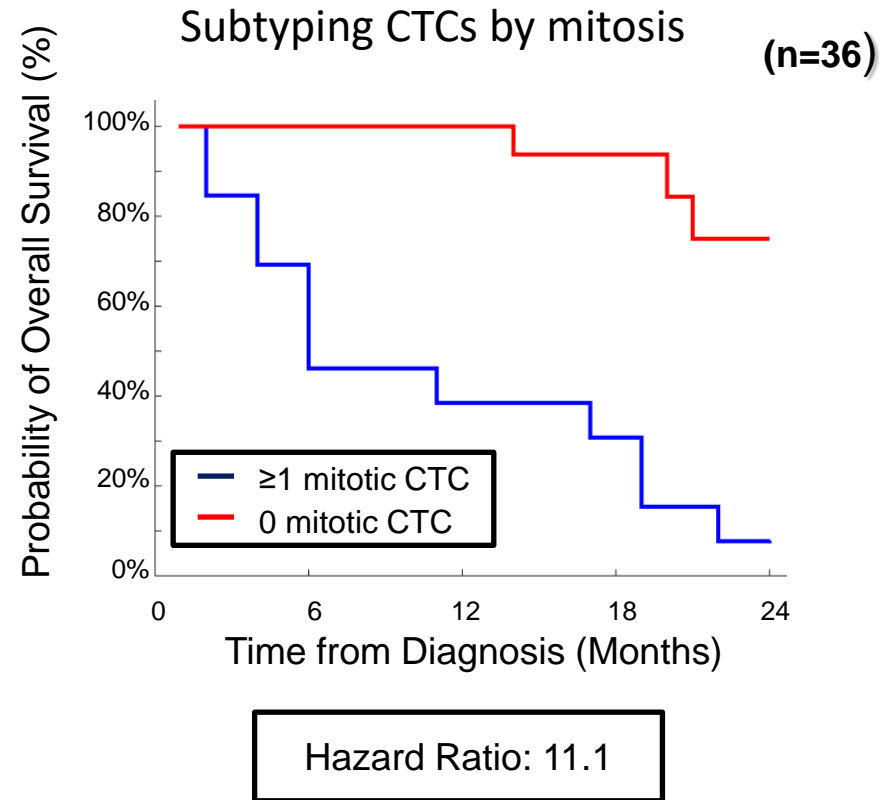
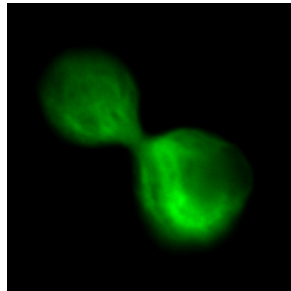
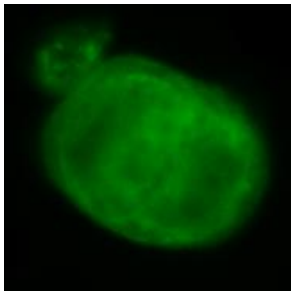
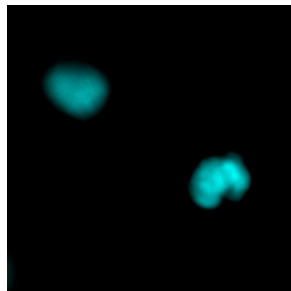
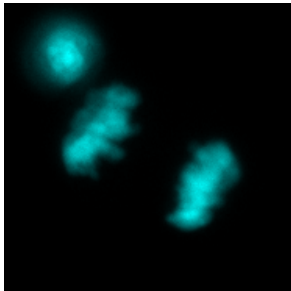
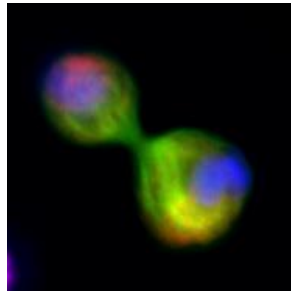
CTCs in Division

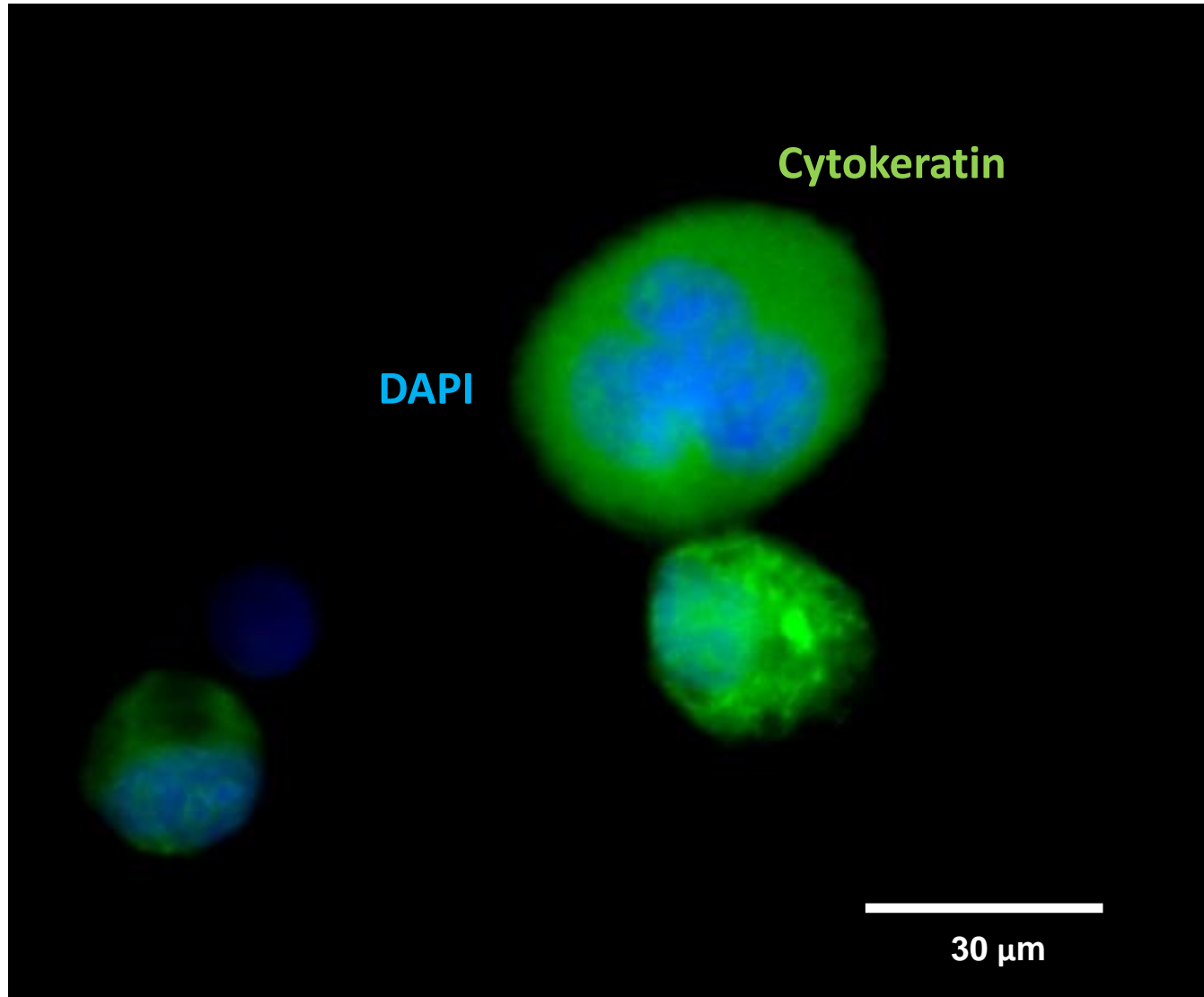
Cytokeratin (green), DAPI(blue)

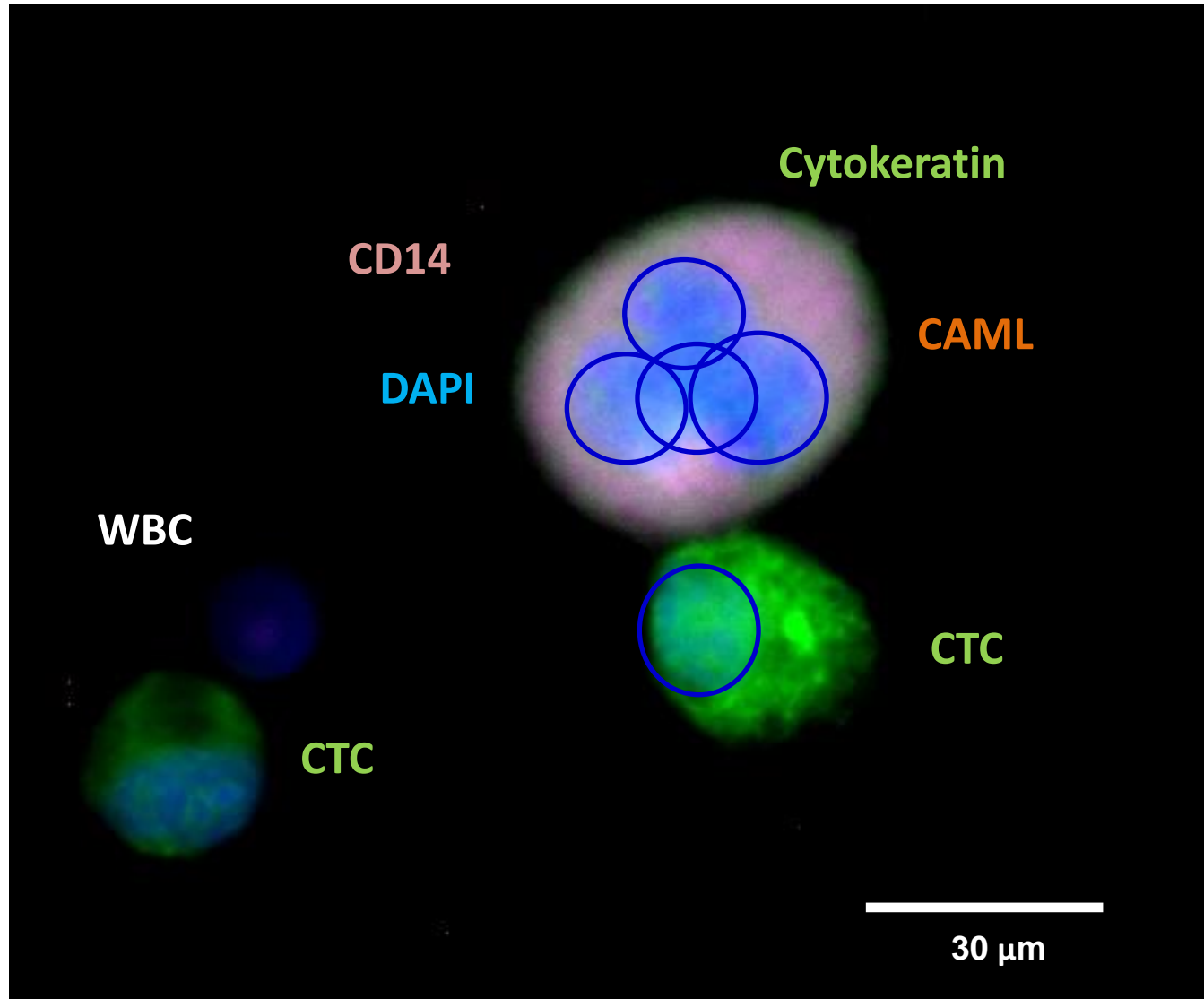
Anaphase



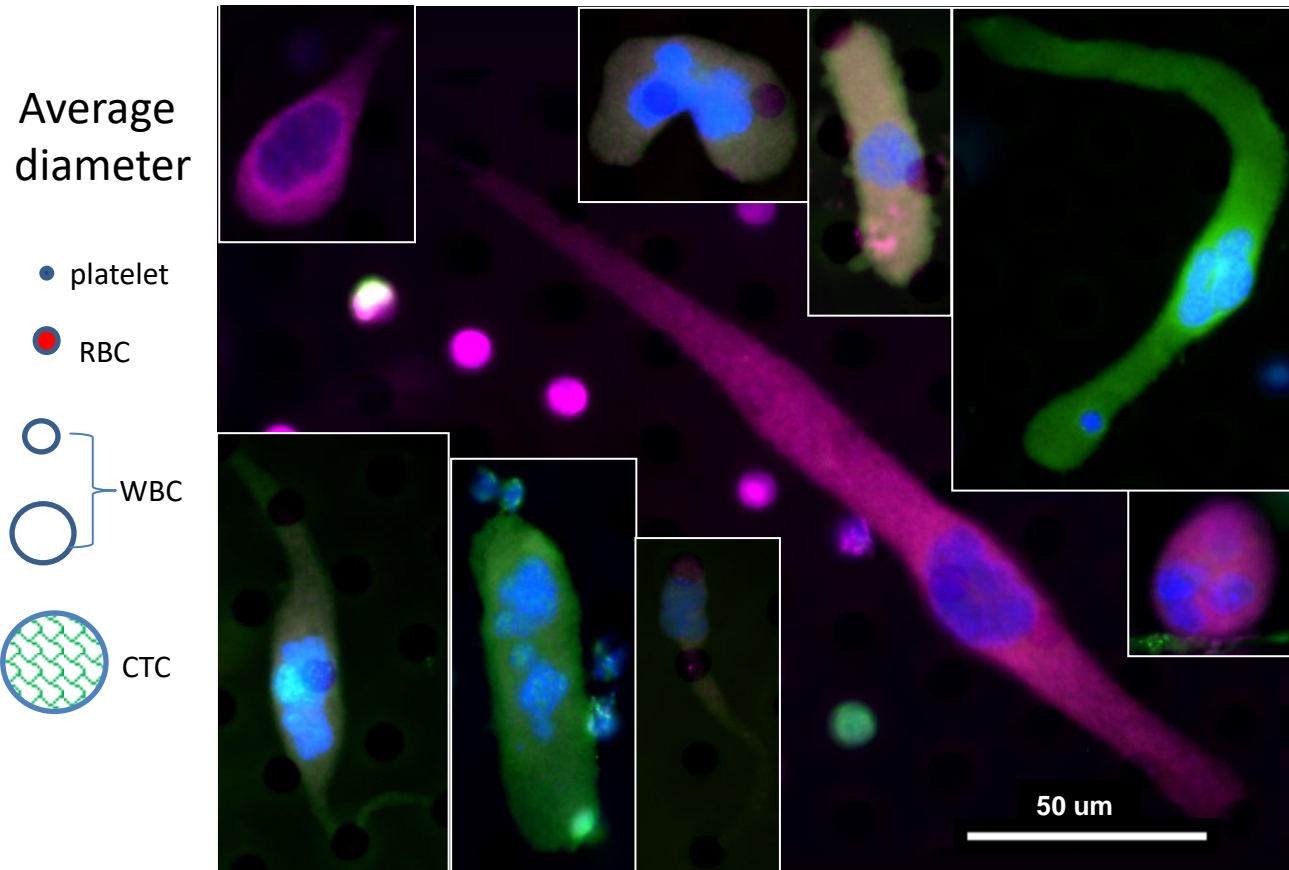
Cytokinesis







Circulating Cancer Associated Macrophage-like Cells (CAMLs)

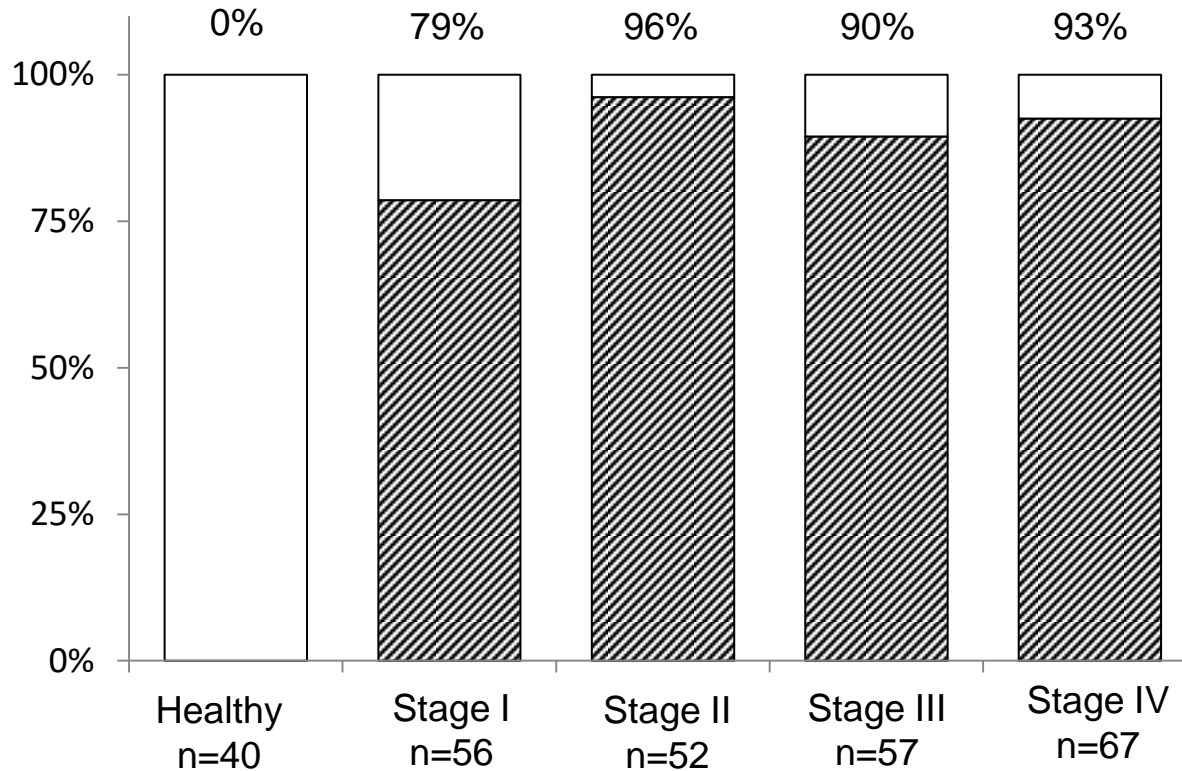


- Large, atypical nucleus
- May express CK and EpCAM
- Contain tumor markers
- Most are CD45 positive
- Large: 30 - 300 μ m
- Express CD11c/CD14
- Express endothelial markers CD146, TIE-2

CAMLs in Cancer Patients

None in healthy controls

Total
n=272



Cancer types

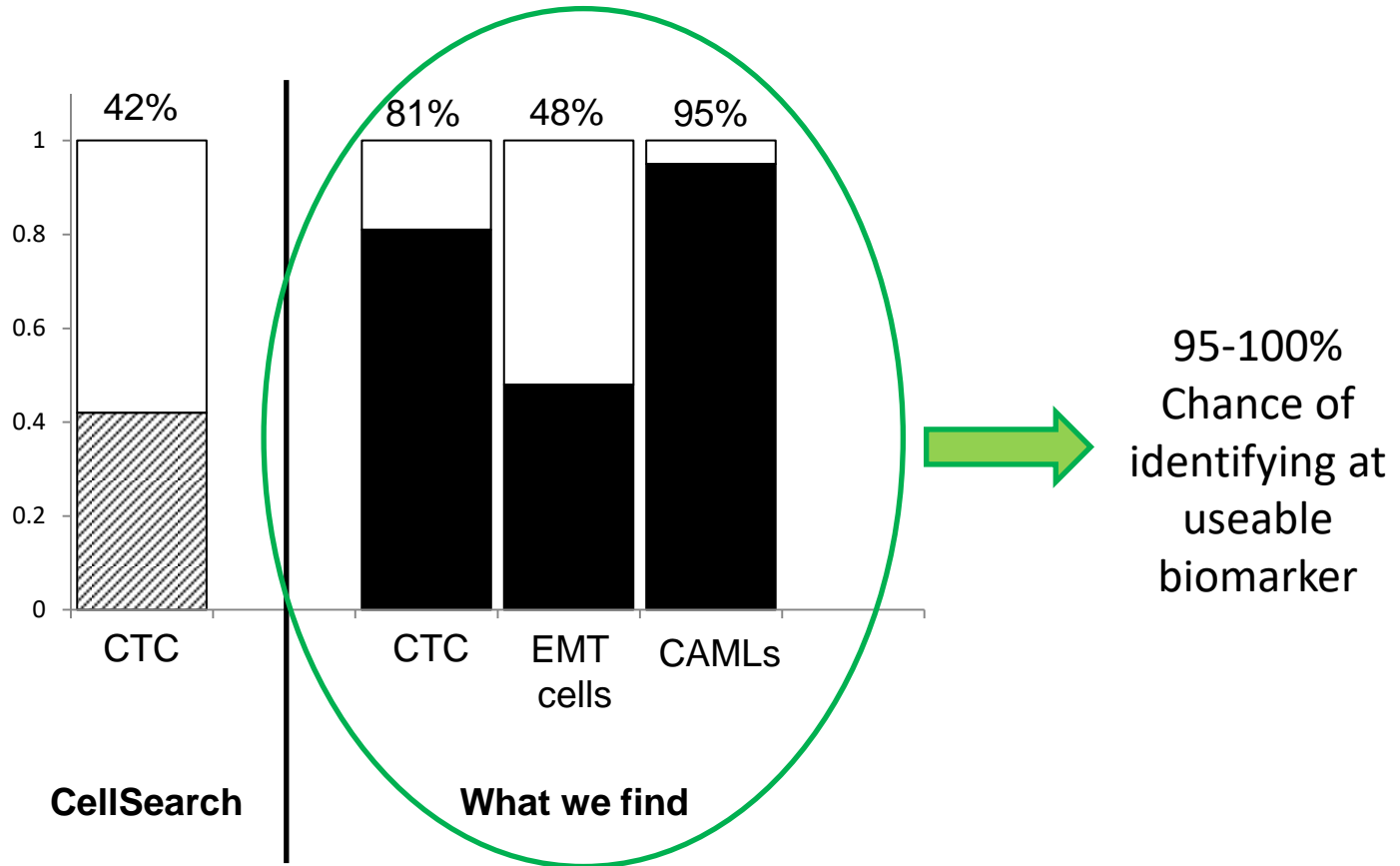
- Breast
- Prostate
- Pancreatic
- Lung (NSCLC)
- Colon
- Esophageal

Sensitivity 89% (95% CI 85-93%)
Specificity 100% (95% CI 91-100%)
PPV 100% (95% CI 98-100%)

Adams, et al. ASCO 2015
 Adams et al CEBP 2016

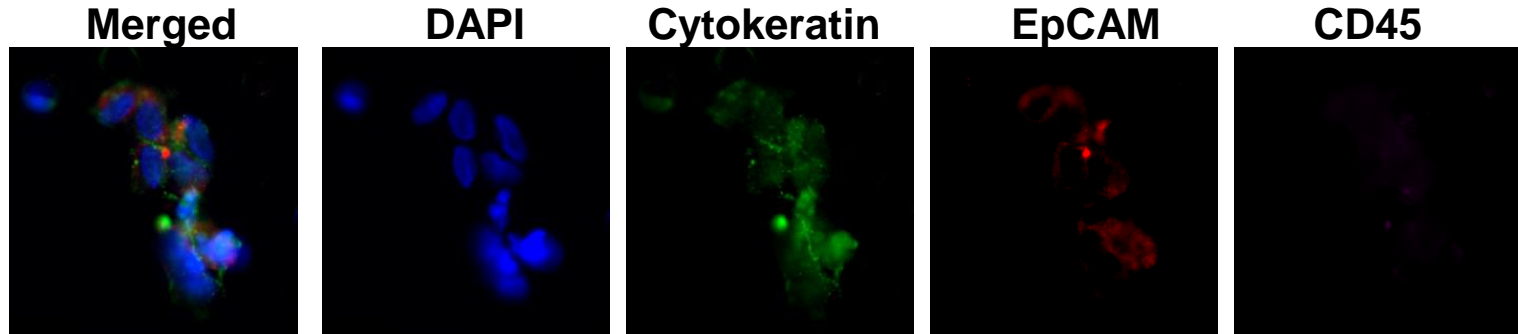
We analyze CTCs and CStCs to maximize useable biomarkers

Presence of cell types in Breast Cancer Patients

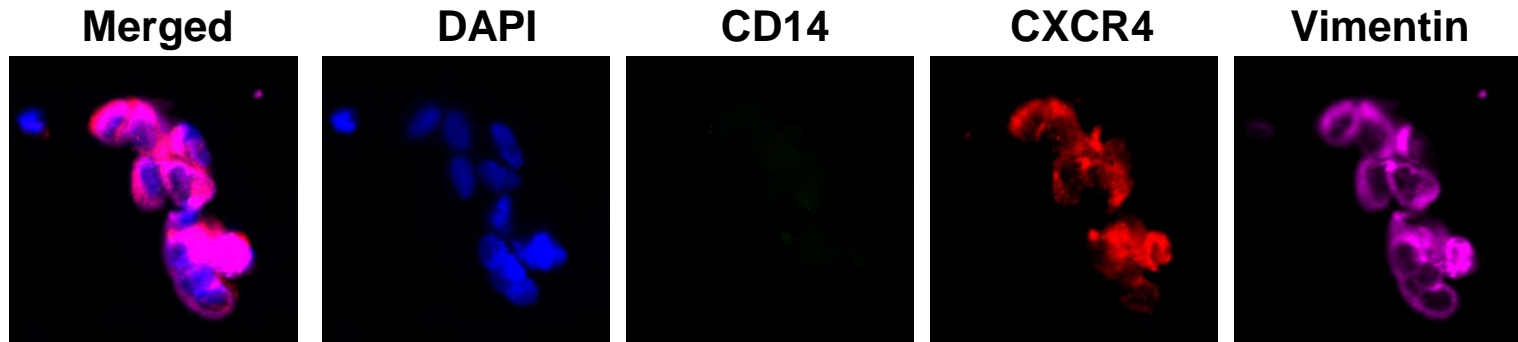


Multi-analyte Subtyping CTCs and CStCs using a single sample

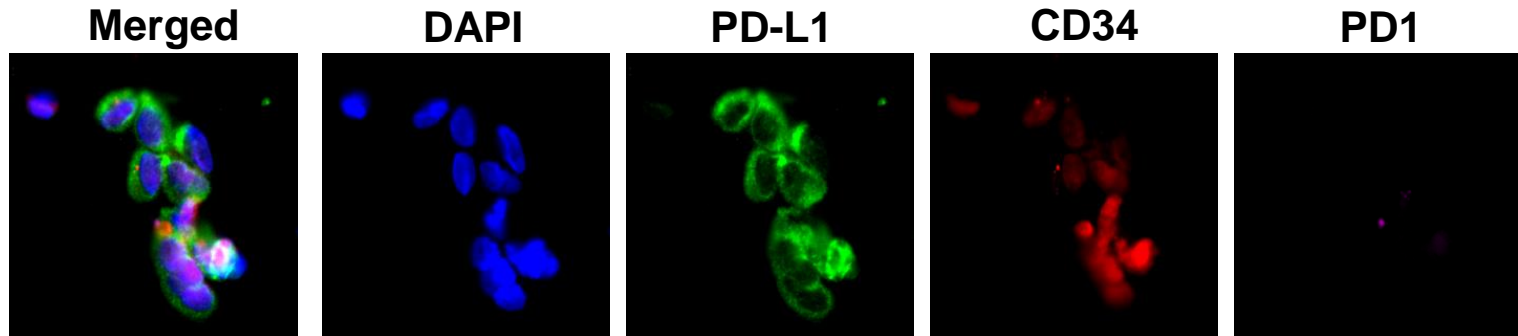
Initial CTC
stains



Subtyping
cells



Therapy
targeting

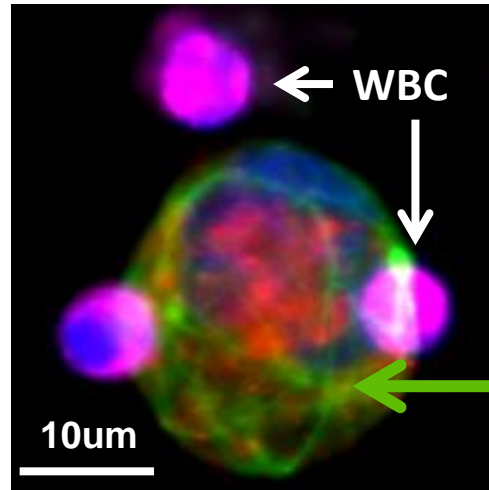


Analysis of Immunotherapy

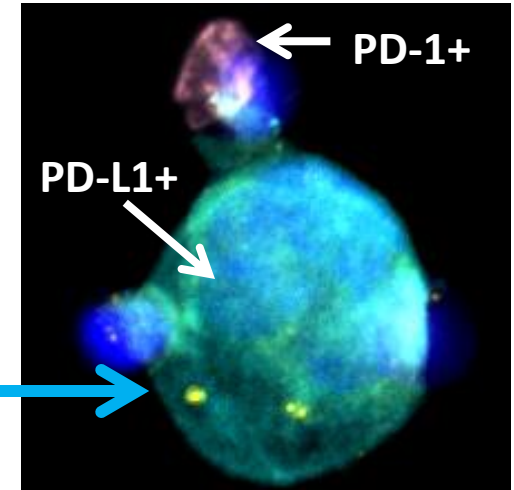
Breast

CTC with bound
white blood cells

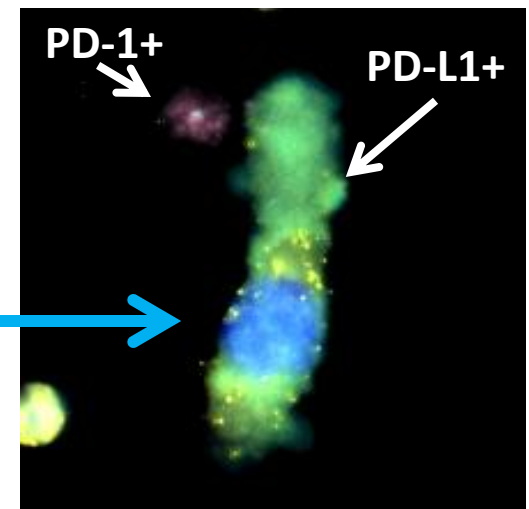
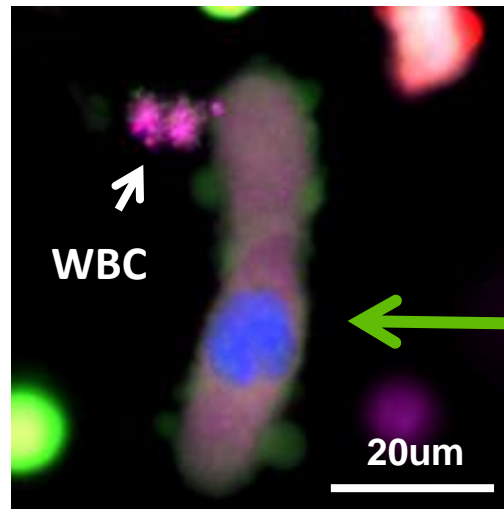
Nucleus(dark blue)/CK(green)/
EpCAM(red)/CD45(violet)



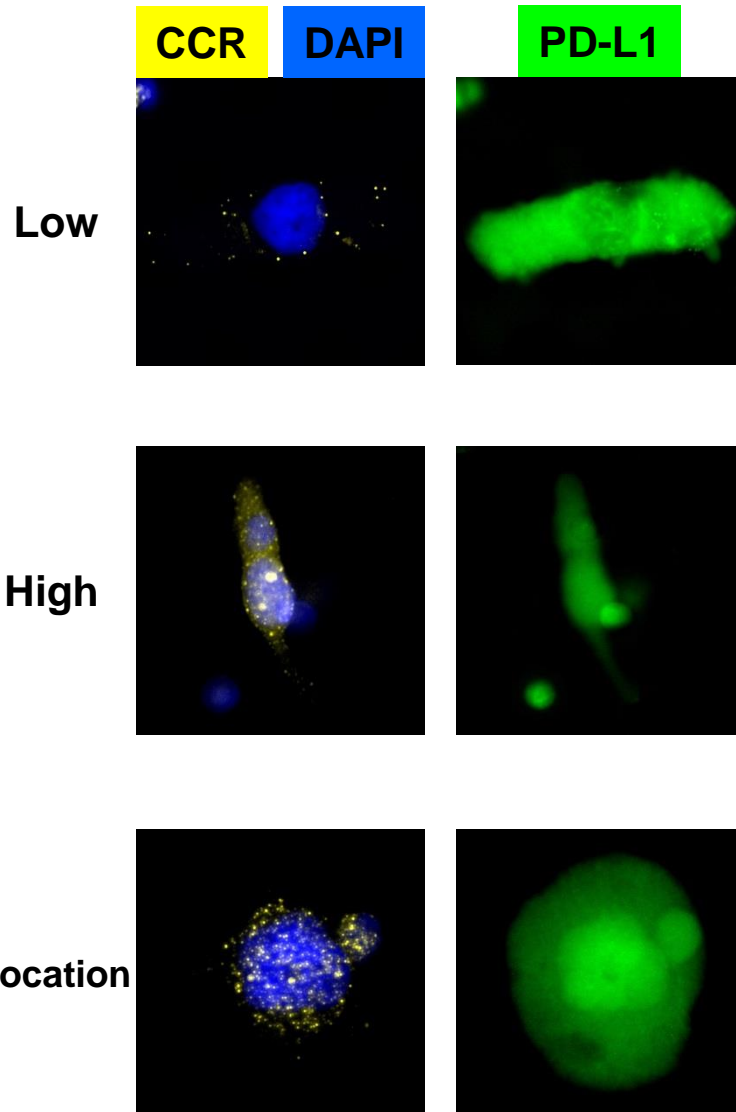
Nucleus(dark blue)/PD-L1(turquoise)/
PD-1(pink)



Cytokeratin positive
CStC with bound
white blood cell



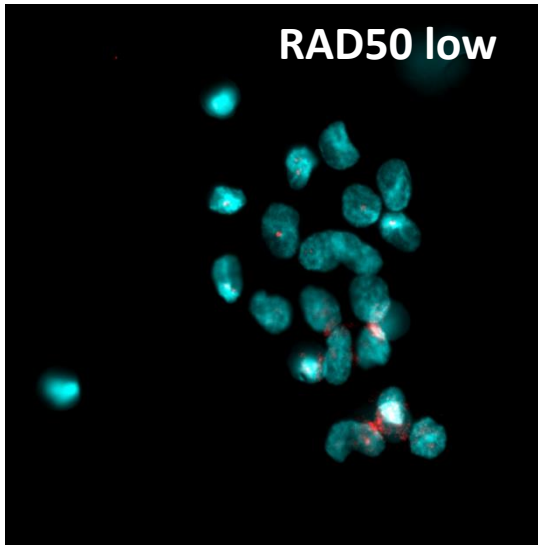
Proteomic profiling of each individual patient in Real-Time



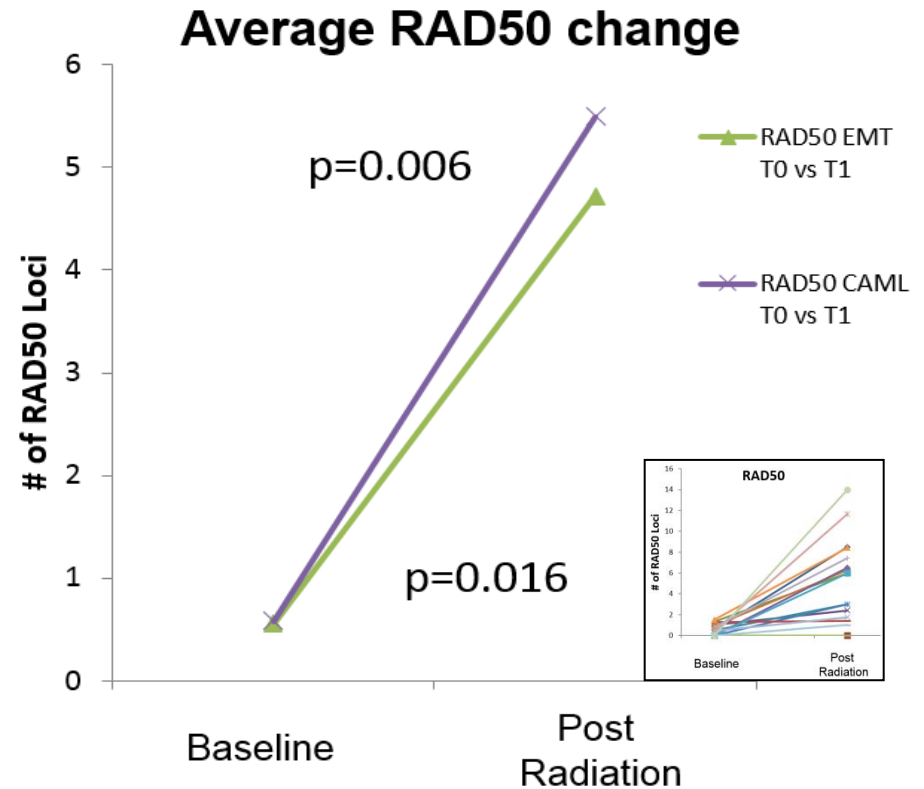
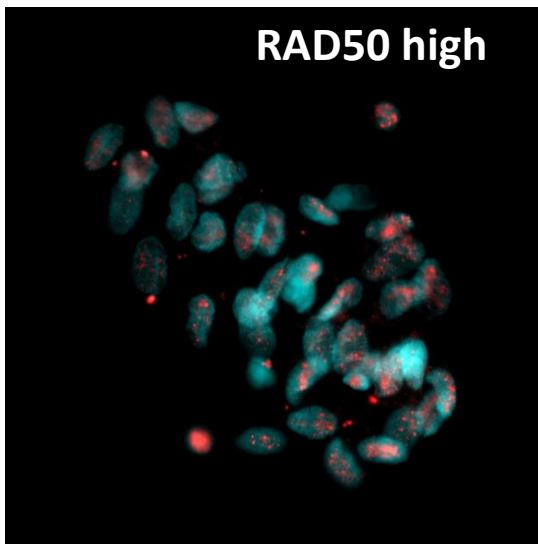
	CK	VM	PD-L1	CXCR4	CD34	EpCAM	CD45	PD-1	CD14
P4									
P6									
P2									
P12									
P11									
P10									
P1									
P3									
P5									
P7									
P9									
P8									

Tracking Origin of CStCs

Low
Before
Radiation



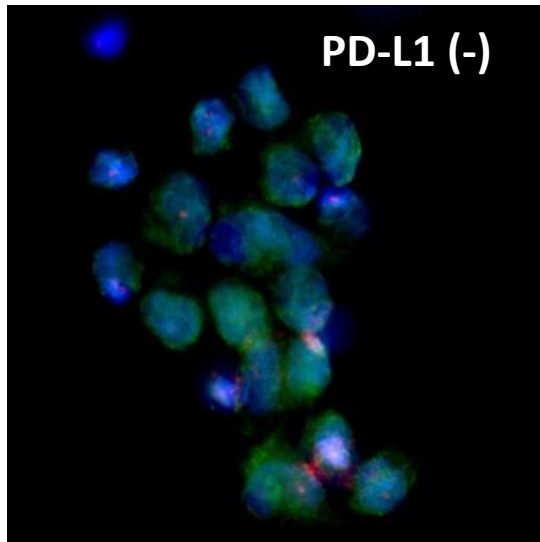
High
Post
Radiation



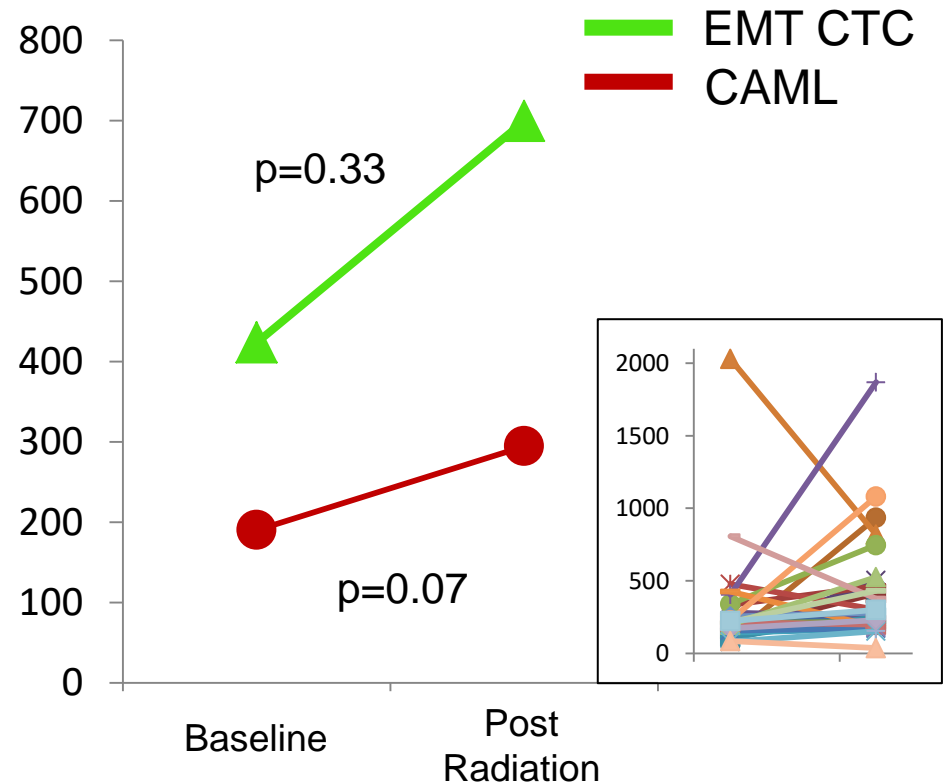
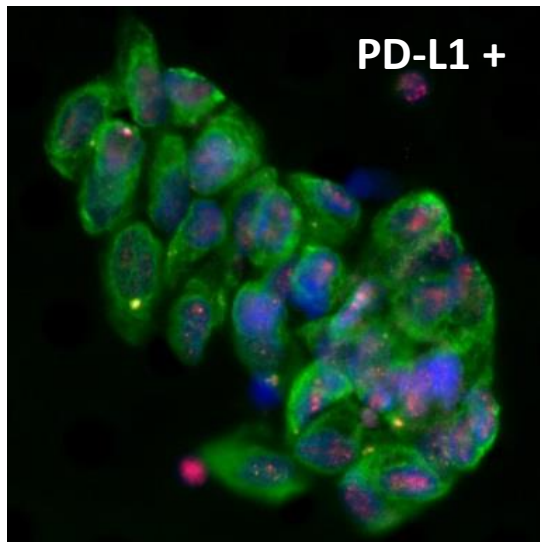
RAD50 foci ranged from 0-20 per cell, with an average of 0.57 at T0 that increased to 5.11 at T1 ($p < 0.001$) during radiotherapy

Tracking upregulation and down regulation of biomarkers in real time

Low
Before
Radiation

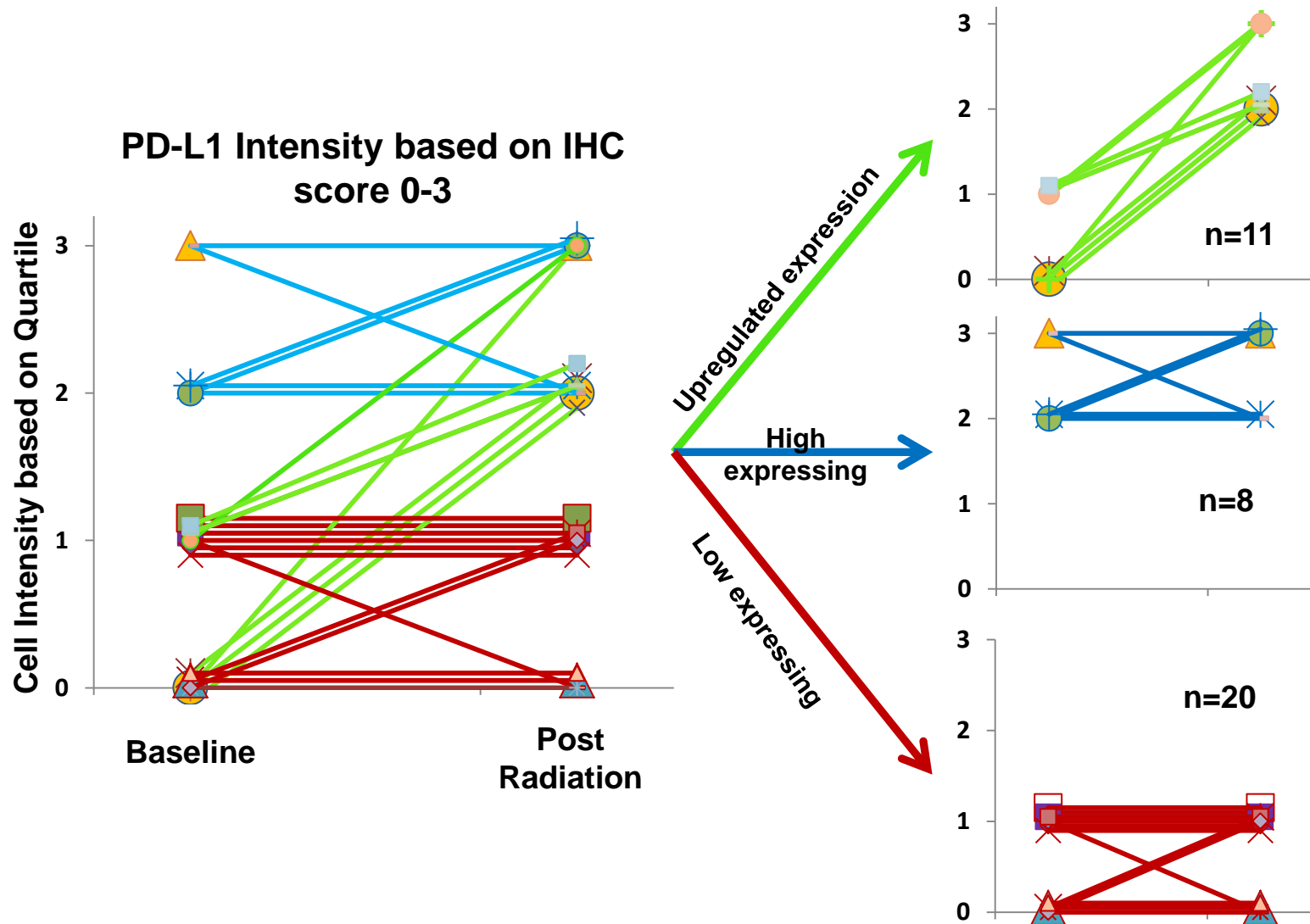


High
Post
Radiation

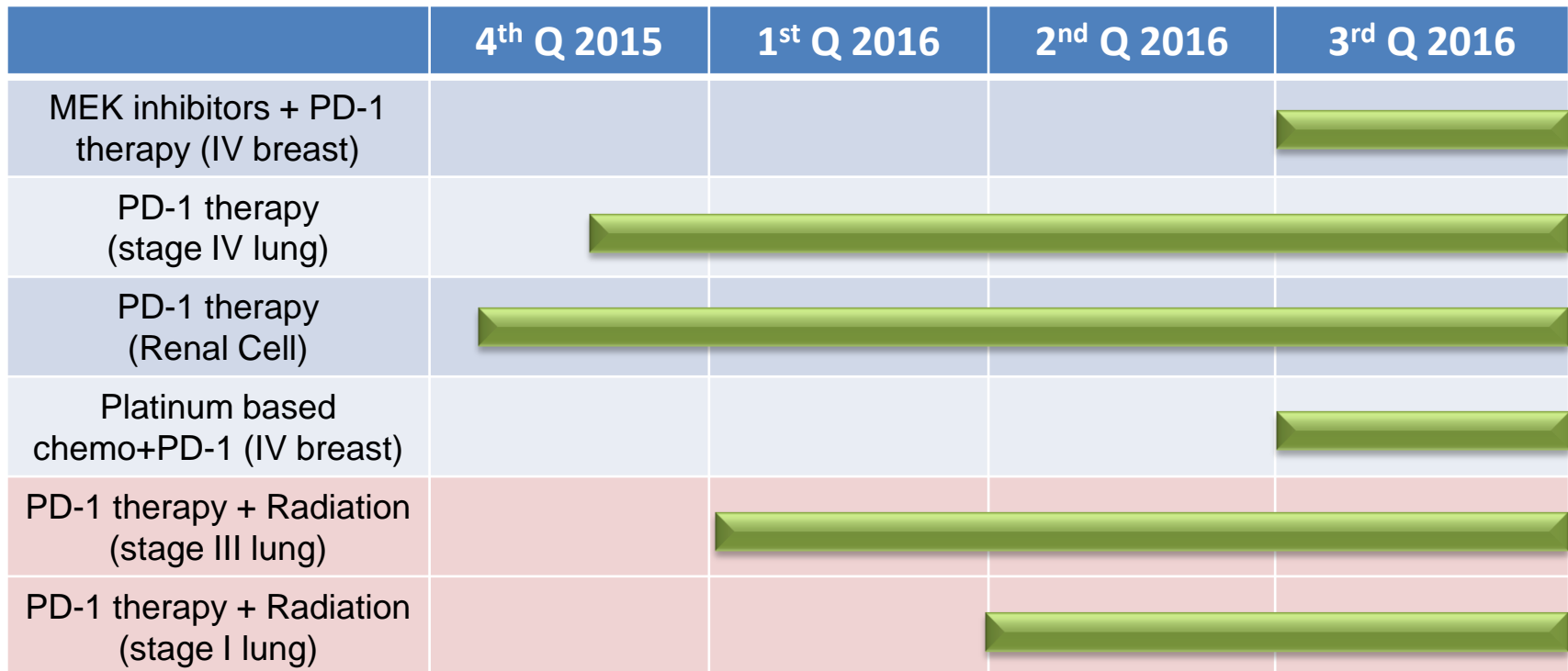


PD-L1 expression ranged from 34-2711 pixel intensity, with an average of 281 at T0 and 565 at T1 ($p=0.07$).


PD-L1 changes in NSCLC patients before and after radiation treatment



Ongoing Research and Clinical Trials with Creatv Partners



 Research Trial

 Clinical Trial

All trials are tracking CTCs, EMTs, and CAMLs

Summary of Creatv's Capabilities

- **CellSieve™ Blood Based Biopsy – isolates CTCs and CStCs**
 - Provides sequential tracking of cancer-baseline through treatment
 - Applicable to ALL Stages of cancer (screening through prognosis)
 - Allows genomic and proteomic profiling of multiple cell types
 - Useful in most solid tumors
- **Analyze all subpopulations of tumor cells and stromal cells**
 - Not marker specific, i.e. multicellular analysis
 - Parallel subtyping of cells and drug targets
- **Wide variety of cancer specific cell analyzes (CTCs and CStCs)**
 - Companion diagnostics
 - Monitor treatment
 - Cancer screening



Research Collaborators

Research Institute	Collaborators
University of Maryland Baltimore	Stuart Martin, Ph.D., Monica Charpentier, M.D. Martin Edelman, M.D., Rena Lepidus, Ph.D.
Northwestern University	Massimo Cristofanilli, M.D.
Fox Chase Cancer Center	R. Katherine Alpaugh, Ph.D.
Johns Hopkins University	David Loeb, M.D.
Mayo Clinic Cancer Center	Thai Ho, M.D., Saranya Chumsri, M.D.
MD Anderson	Steven Lin, M.D.
Medical College of Wisconsin	Susan Tsai, M.D.
OHSU Knight Cancer Institute	Raymond C. Bergan, M.D.
Duke University	Jeffery Marks, Ph.D.
Memorial Sloan Kettering Cancer Center	Daniel Danila, M.D.
Washington University	Rebecca Aft, M.D.
University of Chicago	Susan Cohn, M.D.
George Washington University	Christian C. Haudenschild, M.D.
Hememics Biotechnologies	Steigrimur Stefansson, Ph.D.

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Thank you

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