ASCO, Chicago, IL, May 31- June 4, 2013 ABSTRACT #111046

Low Cytokeratin- and Low EpCAM-Expressing Circulating Tumor Cells in Pancreatic Cancer

Daniel Adams¹, Susan Tsai², Olga Makarova³, Peixuan Zhu¹, Shuhong Li¹, Platte Amstutz⁴, Cha-Mei Tang⁴ ¹Creatv MicroTech, Inc., Rockville, MD, ²Medical College of Wisconsin, Milwaukee, WI, ³Creatv MicroTech, Inc., Chicago, IL, ⁴ Creatv MicroTech, Inc., Potomac, MD

ABSTRACT

To date, circulating tumor cells (CTCs) in the peripheral blood of pancreatic patients using standard immunocapture techniques has met with limited success. As pancreatic cancer is prone to metastasize at distant sites, and therefore should have high number of CTCs, it is possible that immuno-capture methods are not suitable for this disease. Microfiltration is an increasingly popular method for isolating CTCs from the peripheral blood of cancer patients with solid tumors, regardless of surface marker expression²⁻⁴. Using a microfiltration approach, we show that two distinct CTC subtypes can be identified in the peripheral blood in 80% of pancreatic patients.



CellSieve[™] microfilters are lithographically fabricated membranes with high porosity, precise pore dimensions, and patterned pore distribution. We previously reported that CellSieve[™] rapidly and efficiently isolates CTCs from whole peripheral blood, using fluorescent antibody stain as the detection platform. In addition to enumerating CTCs, subtyping by phenotypic determinates may aid in identifying the CTCs cellular status for diagnosis, prognosis and therapy determination.¹⁻⁴

Figure 2. EMT-Like CTCs

- (a) Pancreatic CTC with high EpCAM expression and weak cytokeratin staining
- (b) Pancreatic CTCs with low EpCAM and weak cytokeratin staining

- Neither cell type was found in any healthy subjects (n=30).
 - Merged
- DAPI
- CK 8, 18, 19



MATERIALS & METHODS

Pancreatic patient samples (n=50) were provided by Medical College of Wisconsin, Milwaukee, WI. CellSieve[™] microfilters, with precision 7 micron diameter pores distributed in uniform arrays were employed. 7.5 mL of whole blood was diluted in pre-fixation solution and filtered through CellSieve[™] microfilters (~3 min). CTCs collected were fixed, permeabilized, and stained with DAPI, and an antibody cocktail against cytokeratin (CK) 8, 18 and 19 (FITC), EpCAM (PE), and CD45 (Cy5). CTCs, defined as cytokeratin positive and CD45 negative, were found in two distinct subtypes. One subtype had the "classic" characteristics of a CTC, with high EpCAM and cytokeratin expression, identifiable filamentation, and a cancer-like nuclear structure. The second subtype is





- <u>Oncol.</u> 28 (8):1208-1215
- Pathology 156(1): 57-63.
- 339:580-584

exploited for personalized treatment of cancer patients.

References

Pachmann, K., et al. (2008). "Monitoring the response of Circulating Epithelial Tumor Cells to Adjuvant Chemotherapy in Breast Cancer Allows Detection of Patients at Risk of Early Relapse." J. of Clin.

2. Vona, G, et al. (2000). "Isolation by Size of Epithelial Tumor Cells A New Method for the Immunomorphological and Molecular Characterization of Circulating Tumor Cells." American Journal of

3. Lecharpentier, et al. (2011). "Detection of circulating tumour cells with hybrid (epithelial/mesenchymal) phenotype in patients with metastatic non-small cell lung carcinoma." <u>BJC</u>. 105:1338-1341

4. Yu, M, et al. (2013). "Circulating Breast Tumor Cells Exhibit Dynamic Changes in Epithelial and Mesenchymal Composition." Science