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ABSTRACT

Microfiltration is an increasingly popular method for isolating circulating tumor cells (CTCs) from the peripheral blood of cancer patients with solid tumors²⁻⁴. The microfiltration approach can be used on peripheral blood as a noninvasive "liquid biopsy" for precision cancer detection, regardless of surface marker expression²⁻⁴. Here we describe the use of CellSieveTM microfilters to isolate and subtype CTCs from the peripheral blood of breast, and prostate cancer patients. As it is accepted that CTCs isolated from patient samples represent a highly heterogeneous population with varying degrees of epithelial/mesenchymal differentiation, microfilter isolation may be optimal for the purification of all CTC subtypes. We hypothesize that CTCs from these two different epithelial malignancies can be identified and grouped into distinct subtypes by morphological characterization.



Figure 1. Merged images used dark blue for DAPI. Nuclei are cancerous looking. **Morphological variations of CTCs**

- a) Breast CTC with high EpCAM expression and filamentous CK b) Breast CTC with no EpCAM expression and filamentous CK
- c) Prostate CTC with low EpCAM and very fine filamentous CK

INTRODUCTION

CTCs are cells that originate from a primary solid tumor and are found transiting the circulatory system. It has been well established that CTC enumeration can be used to monitor therapy response and predict outcome.¹⁻ ⁴ Size exclusion is a technique for isolating CTCs from patient samples, irrespective of their surface marker expression.²⁻⁴

CellSieve[™] microfilters are lithographically fabricated membranes with high porosity, precise pore dimensions, and patterned pore distribution.⁵ We previously reported that CellSieve[™] rapidly and efficiently isolates CTCs from whole peripheral blood, using fluorescent antibody stain as the detection platform. In addition to enumerating CTCs, subtyping by phenotypic determinates may aid in identifying the CTCs cellular status for diagnosis, prognosis and therapy determination.¹⁻⁴

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Isolation and Identification of Circulating Tumor Cells (CTCs) from breast and prostate cancer patients





with low expressing EpCAM



Prostate, EMT-Like CTC (c) Prostate, EMT-like cluster of CTCs

MATERIALS & METHODS

Blood from breast and prostate cancer patients were provided by Northwestern University, Fox Chase Cancer Center, and University Maryland Greenebaum Cancer Center, and analyzed by Creatv MicroTech. Microfilters are fabricated with 7 micron diameter pores and uniform array of 160,000 pores over a 9 mm diameter area. 7.5 mL of whole blood was mildly pre-fixed and filtered through CellSieve[™] microfilters (~3) min). CTCs collected were then fixed, permeabilized, and stained with DAPI, and antibodies to cytokeratin (CK) 8, 18 and 19 (FITC), EpCAM (PE), PSMA (Dylight 594) and CD45 (Cy5). CTCs were classified by their morphology, nuclear profile and the expression patterns of cytokeratin, PSMA and EpCAM.

Figure 3. Cluster of CTCs with filamentous cytokeratin from a breast cancer patient

Figure 4. Subtypes of CTCs within cancer families (a) Prostate, early apoptotic, (b)

expression

20 um

RESULTS

The three malignancies have distinct identifiable morphologies Breast – high CK expression in a filamentous pattern (Figs.

Prostate – express PSMA and fine filamentous CK (Fig 1c

Within each cancer CTCs could be subdivided

EMT-like CTCs – low expressing CK with smooth nuclear profile (Fig 4b and 4c)

Apoptotic CTC – spotted nuclear and CK patterns (Fig 4a) CTC Clusters – CTCs found in clusters (Fig 1b, 3, & 4c)

PSMA can verify identified CTCs as prostate cells (Fig. 2).

Phenotypic traits can be identified and classified for comparative analysis (Figs. 1-3).

CONCLUSIONS

Microfiltration captures CTCs regardless of surface marker

CTCs have multiple distinct phenotypes

CTC phenotypes differ between malignant diseases

Microfiltration captures weakened and apoptotic CTCs.

CTC subtypes may indicate definable traits which may be exploited for personalized treatment of cancer patients

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